

Novel and emerging therapies in the treatment of recessive dystrophic epidermolysis bullosa

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Summary

Epidermolysis bullosa (EB) is a clinically and genetically heterogeneous group of inherited blistering diseases that affects ~ 500,000 people worldwide. Clinically, individuals with EB have fragile skin and are susceptible to blistering following minimal trauma, with mucous membrane and other organ involvement in some subtypes. Within the spectrum of EB, ~ 5% of affected individuals have the clinically more severe recessive dystrophic (RDEB) variant with a prevalence of 8 per one million of the population. RDEB is caused by loss-of-function mutations in the type VII collagen gene, *COL7A1*, which leads to reduced or absent type VII collagen (C7) and a paucity of structurally effective anchoring fibrils at the dermal-epidermal junction (DEJ). Currently, there is no cure for RDEB, although considerable progress has been made in testing novel treatments including gene therapy (lentiviral and gamma retroviral vectors for *COL7A1* supplementation in keratinocytes and fibroblasts), as well as cell therapy (use of allogeneic fibroblasts, mesenchymal stromal cells (MSCs), and bone marrow transplantation (BMT)). Here, we review current treatment modalities available as well as novel and emerging therapies in the treatment of RDEB. Clinical trials of new translational therapies in RDEB offer hope for improved clinical management of patients as well as generating broader lessons for regenerative medicine that could be applicable to other inherited or acquired abnormalities of wound healing or scarring.

Keywords: Epidermolysis bullosa, treatment, protein, cell, gene

1. Introduction

Epidermolysis bullosa (EB) comprises a phenotypically diverse group of inherited blistering diseases that affect the skin and, in some subtypes, mucous membranes and other organs (1). Clinically, individuals with EB have fragile skin and are susceptible to blistering following minimal trauma. Depending on the level of blistering within the dermal-epidermal basement membrane zone, EB is classified into four main categories; simplex, junctional, dystrophic and Kindler syndrome (1). The sub-classification of EB extends to over 30 clinical subtypes with pathogenic mutations in at least 18 distinct

genes (2). Within the spectrum of EB, ~ 5% of affected individuals have the clinically more severe recessive dystrophic (RDEB) variant. Dystrophic EB is caused by mutations in the *COL7A1* gene encoding type VII collagen (C7) the major component of anchoring fibril adhesion structures that link the epidermal basement membrane to the subjacent dermis (3,4). Inheritance of DEB can be autosomal dominant (DDEB) or autosomal recessive (RDEB) and all cases result from *COL7A1* mutations; more than 1,500 mutations have been reported globally, most of which are specific to individual families (5). In RDEB, the *COL7A1* pathology usually involves bi-allelic loss-of-function mutations with point mutations or small insertions/deletions leading to nonsense, splice site, frameshift, or occasionally missense mutations disrupting C7 synthesis, secretion and polymerisation and thereby causing structurally defective anchoring fibrils leading to skin fragility. The most severe forms of RDEB are associated with a complete absence of expression of C7 in skin basement membrane leading to no discernible anchoring fibrils (6). In this review, we

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asses novel and emerging therapies in the treatment of RDEB.

2. Current management: Symptoms and complications

The management of RDEB remains complex with no curative therapy currently available. The main principle of care is to manage blisters and erosions, control infection and prevent complications. Symptom relief is very important as both pain and itch have severely deleterious impacts on quality of life. In RDEB, blisters form following minor trauma and/or friction. These blisters need to be lanced to prevent extension of the blister and further skin damage. Pain is a common and constant feature seen in patients with RDEB and arises from four major sources: skin, pain associated with procedures, bone and gastrointestinal (7). For skin care, semi-occlusive dressings that are non-adhesive such as silicone and foam dressings are preferable for treating erosions and reducing skin pain as they absorb exudate and offer some physical protection, thereby providing a moist, clean barrier against bacteria (8). Opioids in the form of morphine, oxycodone, codeine and fentanyl given by a variety of routes including oral, subcutaneous and sublingual are an effective method of relieving most types of pain in RDEB (9). For oesophageal pain, H2 blockers and proton pump inhibitors for gastro-oesophageal reflux can be used and systemic steroids can be utilised during episodes of acute oesophageal blistering (10). Tricyclic antidepressants such as amitriptyline and doxepin taken orally have anecdotally been shown to be beneficial to manage pain in junctional EB (11). Pruritus is a common problem and often correlates with the severity of EB, with RDEB individuals often experiencing significant skin itching (12). The primary cause of pruritus in RDEB remains unclear but has been postulated that wound healing and inflammation may contribute to an itch-scratch-blister cycle leading to further skin damage (13). Menthol containing, oil-based products may be partially helpful in relieving itch (see www.debra-international.org for best practice guidelines).

Oral care is difficult in RDEB due to microstomia, ankyloglossia and vestibule obliteration and so there is a tendency to develop dental abscesses and periodontal disease, both of which can cause pain (14). Caries in RDEB can be reduced through regular dental follow up to optimise oral hygiene and professional cleaning with fluoride therapy (15). Extraction of teeth was previously considered the mainstay of treatment (16) but today prevention of dental disease is the main aim with dentists working closely as part of a multidisciplinary approach (17). Oral pain can be minimised by rinsing the mouth with coating products such as sucralfate or with the use of topical anaesthetics (18).

Insensible losses and thermal dysregulation from chronic wounds leads to a hypercatabolic inflammatory

state requiring an increased calorie intake (19). As a result, the severity of EB often correlates with malnutrition and so RDEB patients often have an inadequate nutrition with growth retardation commonly seen in at least half of all children with RDEB (19). One consequence of inadequate nutrition is pubertal delay and short stature. In most patients with RDEB, bone mineral density is reduced due to poor nutritional status, low 25-[OH] vitamin D levels and reduced mobility (20). In RDEB, bone mineral density and serum bone profile should be monitored and managed with the use of calcium and vitamin D supplements and bisphosphonates to reduce the risk of fractures (21). If pubertal delay is present in RDEB, it is important to attain age appropriate secondary sexual characteristics for psychological reasons and to optimise growth and acquiring peak bone mineral content, therefore, hormonal induction of puberty is often recommended (22).

3. Infection control

Extensive areas of denuded skin pose a risk of skin infection due to the accumulation of serum and moisture that enhances the accumulation of bacteria. Prevention and management of infection is important, as wounds that are chronically colonised heal poorly and slowly (23). In critically colonised wounds, the bacterial load can be reduced with topical agents such as diluted bleach baths, topical antiseptics and topical antibiotics (24). Wounds showing clinical evidence of frank infection require administration of systemic antibiotics with the choice based on culture and sensitivity results.

4. Surgery for contractures

Blisters and wounds in RDEB heal with scarring. This scarring leads to contractures and is most notable on the hands and feet (25). The changes affecting the hands include flexion contractures of the interphalangeal joints, metacarpophalangeal, and wrist joints. In severe forms of RDEB a "mitten" deformity develops with epidermal "cocooning" that encases the hand (26). With minor trauma to the hands and feet, ulceration occurs which can be followed by fibrinous adhesions and scarring, destroying the web spaces and progressing to the finger tips leading to pseudosyndactyly. The term pseudosyndactyly is used as the dermis of the adjacent fused digits remains and separates the fused digits. Pseudosyndactyly of the hands and feet starts in childhood and is characteristic of severe forms of RDEB (27). The formation of scar tissue and contractures causes pain when extending the affected joints (7,9). As dermis abuts dermis in the fused digits, surgery releasing the contractures can exploit this level of fusion, although finding a distinct plane of tissue separation can be difficult in older children and adults. Despite the complexity of surgery, intervention is often

successful in releasing the contractures and separating the fingers, although recurrence of pseudosyndactyly typically occurs. Skin grafting is often required and post-surgical splinting to minimise the speed of recurrence is challenging (26,28).

5. Squamous cell carcinoma

The most serious complication associated with RDEB is the development of clinically aggressive squamous cell carcinoma (SCC) often arising in areas of non-healing cutaneous wounds (29). Based on a US nationwide registry of EB patients, the cumulative risk of first SCC development in severe generalised EB is 7.5% by the age of 20 years. This risk increases to 67.8% by the age of 35, 80.2% aged 45 and 90.1% by the age of 55 (29). Approximately 80% of RDEB patients that develop SCC generally die of metastatic disease within 5 years of excision of the primary lesion (29). SCCs in RDEB can be multifocal and multiclonal with multiple primary tumours co-existing in one individual (30).

Following a systematic literature review and expert consensus, recommendations have been made on the management of cutaneous SCC in EB (31). Wide local excision is considered the treatment of choice for EB-associated SCCs. Imaging with a PET-CT scan evaluates distant disease and should underlying vessels, nerves or tendons be involved, then more radical surgery such as amputation may be more appropriate (31). Under circumstances when there has been local recurrence of disease or regional or distant metastasis, non-surgical treatment such as radiotherapy or chemotherapy may be considered. Topical preparations such as photodynamic therapy and 5-fluorouracil have been used in a small number of patients with in-situ disease (31). When using radiotherapy, consideration needs to be given to severe desquamation that can follow larger total radiation doses. Conventional chemotherapy has been used in cases of advanced EB SCCs (32-35). Agents have included cisplatin, carboplatin, paclitaxel, fluorouracil, doxorubicin and methotrexate. Partial remission has been described in some reports although follow up data are limited. Newer biologic agents such as epidermal growth factor receptor (EGFR) antagonists and tyrosine kinase inhibitors have been used in non-EB SCCs (36-38), but reports of their use in EB are few. Cetuximab, a monoclonal antibody that binds the extracellular domain of EGFR has shown favourable results in metastatic EB SCCs strongly expressing EGFR, although numbers of cases are limited and long term survival remains poor (39,40). Systemic retinoids have been trialled in RDEB as a chemopreventative agent to reduce the risk of SCC. A phase 1 trial of isotretinoin in twenty RDEB patients (41) showed no adverse reactions at a low dose of isotretinoin however, increased mechanical fragility was observed at therapeutic doses and so currently, retinoids are not recommended for long term chemoprophylaxis.

6. Wound grafting and topical therapies

A number of biological dressings and wound grafting approaches have been used to treat intractable ulcers in RDEB (42-46). Autologous and allogeneic skin grafting have been developed for RDEB with some reported success, mostly in small case series or anecdotal reports. In one study, cultured epidermal autograft (CEA) was manufactured by taking a full-thickness biopsy specimen of skin from an RDEB subject and culturing keratinocytes to confluence. The resultant CEA was then grafted onto a designated area of ulceration with epithelialisation observed 2 weeks later (42).

Allogeneic cultured dermal substitutes (CDS) have also been used to treat intractable ulcerated wounds in patients with RDEB (44,47). Apligraf® (Organogenesis, Canton, MA, USA) is an allogeneic cultured skin substitute consisting of keratinocytes and fibroblasts supported on a scaffold and was initially used in the treatment of venous ulcers. However, Apligraf® has also been used to treat EB skin ulcers with benefit, although mainly in subtypes of EB other than RDEB (43,44).

CDS have been used in several patients with RDEB with reported success (45,46) although long term improvements may be limited and repeated preparation and application of skin grafts may not be practicable or economically feasible.

Alternatively, amniotic membrane, which possesses biological properties that can promote wound healing (48), has been used in EB to promote healing of chronic wounds (49). In a retrospective, proof-of-concept study, amniotic membrane grafting was efficacious in promoting the healing of non-healing wounds in EB with a reduction in pain but complete re-epithelialisation was not achieved (49). An additional study in DEB examined clinical application of amniotic membranes if the wound was debrided and found there was spontaneous reepithelialisation in a week and pain and immobility improved within hours (50).

Placental material has also been used to manage acute and chronic wounds. Cryopreserved placental membrane (CPM) (Grafix, Osiris Therapeutics, Inc., Columbia, Md.) is a cellular matrix composed of placental membrane matrix that provides the wound with mesenchymal stem cells, neonatal fibroblasts, epithelial cells, growth factors (GFs), and angiogenic factors and has been licensed for the management of EB (51,52). Although trial data for RDEB are lacking, CPM showed superior results to standard wound care in a randomised controlled trial comparing the two treatment modalities to treat diabetic foot ulcers, and finding that wound closure at 12 weeks was significantly higher in the CPM group (62% in the CPM arm vs 21% when standard wound care was used) (53).

Acellular dressings with collagen derived from a variety of sources have also been utilised to improve wound healing in RDEB (54). The rationale for their use,

includes observations that type I collagen may decrease MMP activity and act as an anti-inflammatory agent by binding pro-inflammatory cytokines (55). Integra® (Integra LifeSciences, Plainsboro, NJ) is a bilayer wound dressing with acellular bovine collagen and chondroitin-6-sulphate. Helicoll® (Encoll, Fremont, CA) is a single-layer acellular matrix of purified bovine type 1 collagen and has been trialled in patients with RDEB with the primary outcome being wound size measurement (56), with a statistically significant improvement in wounds treated with Helicoll® compared to standard dressings. However, upon discontinuation of the type 1 collagen treatment, wounds that had re-epithelialised, soon broke down again with recurrent ulceration.

In addition to wound grafting, topical therapies are also being developed to aid wound healing in RDEB such as thymosin β 4, a small molecular weight protein involved in cell proliferation, migration and differentiation, as well as actin polymerisation, which appears to enhance epithelial wound healing when applied topically to wounds in animal studies. The basis of the positive response may involve promoting the migration and adherence of keratinocytes on wounds, and the upregulation of one or more extracellular matrix proteins, particularly laminin-332. A clinical trial to explore the potential of thymosin β 4 to promote wound re-epithelialisation in EB was initiated in 2005; this was a randomised double-blind study involving three concentrations of the agent and a placebo control. However, the study had to be terminated early due to lack of subject recruitment, although no adverse events were reported in those who participated (57).

Topical growth factors have been used in wound healing in venous leg ulcers (58) and diabetic foot ulcers (59). However, a topical preparation of PDGF (platelet-derived growth factor) named Regranex® (Smith and Nephew, London, UK) was trialled in a randomised, placebo controlled, double blind trial which showed no significant improvement in the healing of diabetic foot ulcers (59). Generally, however, the overall efficacy of topical growth factor preparations has been relatively disappointing, and there have been no reported studies in RDEB.

7. Systemic treatment

Before the genetic basis of dystrophic EB was discovered, ultrastructural studies indicated possible collagen degradation and phagocytosis of collagen fibrils in areas of blistering in RDEB skin (60). Thus early attempts at systemic treatment for RDEB focused on inhibiting collagenase. Phenytoin, an anticonvulsant that also has properties as a collagenase activity inhibitor, was trialled in 17 unselected RDEB patients (61). After up to a maximum of 15 months of therapy, blisters and erosions were significantly decreased in most of the patients (61). In 1992, however, a multi-centre

randomised, placebo-controlled, double blind, cross over study of phenytoin in RDEB was performed which showed unequivocally that phenytoin had no significant therapeutic effect (62). Thus, there is currently absolutely no clinical rationale for the ongoing prescribing of phenytoin for the treatment of RDEB.

Following on from the proven failure of phenytoin therapy, but still pursuing the anti-collagenase strategy, minocycline was trialled in two patients with DEB (63), on the basis that tetracyclines (including minocycline) have anti-collagenase activity (64). After commencing minocycline at a dose of either 100mg twice daily or 50mg three times daily blistering was reduced in both subjects (63). Similar benefits have also been reported in a patient with dominant DEB. Regarding mechanism of action, it has been shown that levels of matrix metalloproteinase-9 (MMP-9) are raised in RDEB blisters (65) and it was thought that the clinical improvement might be due to inhibition of MMP-9 by minocycline (66). Nevertheless, minocycline also has a tendency to induce skin hyperpigmentation as a side effect. To date there has been no larger clinical trials to assess clinical use of minocycline in RDEB and thus its use cannot be recommended for routine treatment.

Other antibiotics have been trialled in RDEB, including trimethoprim for its anti-inflammatory effects based on diminished chemotaxis of polymorphonuclear leukocytes, modification of complement pathways and inhibition of MMPs (67). In a proof-of concept double blind randomised cross-over trial comparing trimethoprim to placebo in RDEB, there was a trend towards improved wound healing with trimethoprim compared to placebo (68) although further assessment will be required before trimethoprim might be recommended for routine clinical use. Another preparation that is able to regulate MMP activity *in vitro* and *ex vivo* is the green tea extract, epigallocatechin-3-gallate (EGCG) (69). A multicentre, randomised, crossover, double blind, placebo controlled clinical trial in 17 RDEB individuals evaluated whether a 4 month course of oral EGCG might be efficacious in improving skin impairment (70). Despite the EGCG group having less daily blisters and shorter wound healing times, however, the study failed to demonstrate statistical significance between the two groups. Thus no formal recommendations can be based about the use of oral EGCG in RDEB based on this single study.

Regarding other anti-inflammatory drugs, ciclosporin was discovered to have clinical benefits in the treatment of DEB when prescribed to prevent graft rejection in a child with DEB (71). However, given the increased risk of skin malignancy in RDEB, long term use of ciclosporin cannot be recommended. For other immunosuppressant drugs, a randomised controlled double blinded study in 35 patients with DEB was conducted to evaluate ciclosporin versus mycophenolate mofetil (MMF). The percentage of improvement in the

ciclosporin group was statistically significantly higher than the MMF group but there was no difference in the number of new blisters or the rate of healing of new blisters between the groups (72). As for ciclosporin, however, long term use of MMF in RDEB is not advisable.

In other anecdotal reports, the tumour necrosis factor alpha (TNF- α) inhibitor etanercept has been assessed in RDEB (73). Etanercept is a fusion protein produced by recombinant DNA and is used to treat a variety of disorders mediated by excess TNF- α such as psoriasis and psoriatic arthritis. A 29-year-old woman with concomitant DEB and psoriatic arthritis was given etanercept to treat her psoriatic arthritis. A progressive improvement in her DEB was noted in the first 3 months of treatment with subcutaneous etanercept, 50mg twice a week, with an improvement in pruritus and fewer blisters; notably, the clinical improvement persisted over the 3 years she was receiving etanercept (73).

A patient with RDEB undergoing bone-marrow transplantation (BMT) for her disease (see bone marrow transplantation section) observed that there was a significant improvement in her wound healing during autologous peripheral blood stem cell mobilisation with systemic granulocyte colony-stimulating factor (G-CSF) prior to the transplant (74). Based on this anecdotal finding, a pilot trial was designed to confirm the safety of daily doses of G-CSF, (10 μ g/kg/dose) in 6 RDEB and one DDEB subject. The patients were re-evaluated at Day 7 and for all patients combined, median reductions of 75.5% in wound size and 36.6% in blister/erosion counts were observed. G-CSF was well tolerated and no adverse events were noted. At the request of some individuals, further injections of G-CSF were administered which demonstrated that the response was reproducible and safe (74).

In addition to strategies employed to correct the causative pathology in RDEB, there is also a need to treat collateral pathology such as scarring. The functional limitation of movement secondary to extensive scarring and fibrosis is a major complication of RDEB. A hypomorphic mouse model suggests that this scarring and fibrosis is driven by transforming growth factor beta-1 (TGF- β) signalling, as reflected by transition of dermal fibroblasts to myofibroblasts with capacity for ECM production (75). Losartan, an angiotensin II type 1 receptor antagonist, that is primarily used to treat hypertension, has also been shown to possess anti-fibrotic effects resulting from suppression of TGF- β 1 *via* angiotensin II type 1 receptor mediated down regulation of TGF- β 1 activators such as thrombospondin 1 (TSP-1). TGF- β activity is elevated in injured RDEB skin, and so by targeting TGF- β activity, fibrosis may be reduced and in turn, delay mitten deformity development (75). In murine studies, losartan has been shown to reduce TGF- β levels in RDEB cells *in vitro*, and in the skin and the circulation of RDEB mice. As a result of

reduced TGF- β activity, there was significantly slower progression to fibrotic digit fusion and mitten deformities (76). The role of TGF- β signalling has been highlighted as a potential modifier of disease severity following the study of monozygotic twins with RDEB with markedly different clinical phenotypes and similar amounts of C7 expression (77). In this study, genome wide expression analysis in twins' fibroblasts showed differential expression of the genes associated with TGF- β pathway inhibition. Decorin, a skin matrix component with anti-fibrotic properties was more expressed in the skin of the less severely affected twin. Fibroblasts from the more affected twin were characterised by enhanced α -smooth muscle actin and plasminogen activator inhibitor 1 expression, collagen I release and collagen lattice contraction.

Preclinical studies are also ongoing to evaluate the reparative potential of high mobility group (HMG) proteins, specifically by mobilising key epithelial progenitors from bone marrow which are then recruited to damaged RDEB skin. Murine studies have demonstrated that one of the HMG proteins, high mobility group box-1 (HMGB-1), is rapidly released from hypoxic keratinocytes, such as from blister roofs, and upon release into the circulation, reparative epithelial progenitor cells (Lin-/PDGFR α +) are mobilised from within the MSC-BM population (78). These cells are recruited along a concentration gradient to the area of hypoxic skin damage. Differentiation of these cells into keratinocytes (rather than fusion) was clearly demonstrated, with persistence of the differentiated BM cells in the skin after several renewals of the murine epidermis, data which support engraftment of a murine BM population that has generated keratinocyte stem cells (78).

8. Cell therapies

8.1. Allogeneic fibroblasts

Fibroblasts have the capacity to synthesise C7 as well as modulating wound healing (79). On this basis, a number of RDEB murine and human studies have been conducted injecting allogeneic normal human fibroblasts intradermally with the aim of potentially increasing C7 expression and also improving wound healing (80).

A proof-of-concept study in 5 RDEB individuals demonstrated that a single intradermal injection of allogeneic fibroblasts (5×10^6 cells injected into the superficial dermis over $\sim 1 \text{ cm}^2$) increased *COL7A1* expression for at least 3 months in most subjects (80). The study also demonstrated the low immunogenicity of allogeneic fibroblasts and lack of host response at an immunological and histological level. The injected cells were not detectable at 2 weeks post-injection, the time-point at which an increase in C7 protein at the DEJ was

seen. In murine studies, it has been suggested that this increase in C7 protein at the DEJ may be secondary to donor fibroblasts releasing wild-type full length C7 that can be incorporated into the DEJ for the short time that these donor fibroblasts are present (81). Of note, in the human studies, the increase in C7 was most apparent in RDEB individuals who had some baseline expression of C7 compared to those who had a complete absence of the protein. The source of the new C7 is likely to reflect upregulation of the RDEB subjects' own mutant, but partially functional C7, a mechanism supported by a lack of new normal-appearing anchoring fibrils. A further study showed that a single injection of allogeneic fibroblasts could increase *COL7A1* expression for 3-6 months and C7 protein for 9-12 months (82). The expression of heparin binding-EGF-like growth factor (HB-EGF) was thought to mediate this increase in endogenous C7 expression (82).

With regard to wound healing, a phase II double-blinded, randomised, controlled trial in RDEB patients comparing injections of allogeneic cultured fibroblasts in suspension solution versus suspension solution alone, with the injections given across eroded areas found that in both arms there was a reduction in erosion size, suggesting that perhaps the trauma of either injection might, at least in part, be responsible for the clinical responses (83). On the other hand, a further prospective, randomised, double-blind, within-patient, vehicle-controlled trial of subjects with RDEB was conducted in 11 patients. Twenty-six erosions were treated; 14 with a single treatment of 5×10^6 allogeneic fibroblasts per linear cm of erosion margin and 12 with vehicle. Fibroblast injections produced a greater reduction in erosion area than did vehicle alone during the first 28 days. After 28 days, there was no significant difference between fibroblasts and vehicle although further injections were not administered (84).

8.2. Mesenchymal stromal cells

Multipotent mesenchymal cells are found in several tissues, including the bone marrow (85,86) and have the ability to migrate to injured tissue and stimulate tissue regeneration, thus making this therapy potentially relevant to RDEB wounds. The clinical use of MSCs in RDEB was first reported in a 13-year-old and 25-year-old patient from Chile in 2010 (87). The MSCs were derived from the bone marrow of healthy, unrelated individuals and injected intradermally. Both subjects had clinically severe blistering with a complete absence of C7 expression. Either 0.5×10^6 MSCs or vehicle were injected into both intact and chronically ulcerated sites. At week 12, wounds treated with MSCs had almost healed compared to sites treated with placebo with benefits lasting for 4 months post injection. Thereafter, skin fragility resembled baseline with ulceration. New C7 was seen in a linear pattern at the

junction between the epidermis and dermis, suggesting that intradermal administration of allogeneic MSCs may lead to *de novo* C7 expression in the skin as well as prevention of blistering and improvements in wound healing in patients with RDEB.

Subsequently, El Darouti *et al.* (88) conducted a double-blind study, randomising 14 patients with clinically severe RDEB into two equal groups. Both groups received intravenous MSCs derived from healthy bone marrow aspiration from one healthy parent but group one was also given 5 mg/kg/day of ciclosporin to reduce inflammation or protect against rejection with the patients in group two receiving a placebo suspension. Both groups were seen fortnightly for 12 weeks and were reported to have fewer new blisters, to have an increased rate of wound healing, and to demonstrate new anchoring fibrils on skin biopsies. Two individuals demonstrated clinical benefit at 12 months, whereas the improvements in the remainder peaked 3 months after infusion and waned thereafter.

Petrof *et al.* (89) enrolled 10 children aged 1-11 years in the U.K. with RDEB who had partial or complete absence of C7 protein, in an open-label, phase I/II clinical trial. Each child received three IV infusions of either 20×10^6 cells per infusion (weight ≤ 20 kg) or 40×10^6 cells per infusion (weight > 20 kg) (equivalent to $1-3 \times 10^6$ cells per kg) of BM-MSCs on days 0, 7 and 28. No severe adverse events occurred (other than the transient noxious smell associated with the preservative dimethyl sulphoxide). Skin biopsies revealed no increase in C7 and no new anchoring fibrils at day 60 post infusion. One subject showed no clinical benefit, whereas two had sustained improvement at one year, and in the others there were transient improvements such as less skin redness, less skin pain and itching, and better wound healing that lasted for 4-6 months after the third infusion of MSCs. The optimal dosing, route of administration and consequences of multiple repeat dosing of allogeneic MSCs in RDEB has yet to be fully evaluated. However, murine studies have shown the impact and superiority of high density intradermal injections of MSCs compared to fibroblasts, suggesting that further human clinical trials are needed if the maximal benefits of MSC cell therapy in RDEB are to be realised (90).

The mechanism by which MSCs lead to a clinical improvement in wound healing in RDEB has not yet been established but seems to be indirect and trophic through the release of various growth factors and cytokines (91), *i.e.* without the need for the MSCs to engraft. MSCs express tumour necrosis factor alpha (TNF α)-stimulated protein 6 (TSG-6), which in other studies has been associated with an improvement in wound healing and downregulation of B-cell proliferation, monocyte maturation, secretion of IFN- γ and TNF- α at wounded tissue sites (92), while also promoting increased secretion of anti-inflammatory

IL-10 from macrophages (93). In addition to TSG-6, MSCs also mediate immunosuppression through the secretion of nitric oxide, transforming growth factor-beta (TGF- β) and indoleamine 2,3-dioxygenase (94).

Regarding other cells, potentially with stem rather than stromal functionality, human umbilical cord blood derived unrestricted somatic stem cells (USSCs) have shown potential to regenerate RDEB skin in animal models (95). In murine models, it has been shown that USSCs express C7 and accelerate wound healing when injected intradermally in mice that have full-thickness excisional wounds (96). An intradermal injection of USSCs modified with a luciferase reporter gene, injected at a distant site to the wound revealed specific migration to the wound (96). These data suggest that CB-derived USSCs may contribute to wound repair and may be worth exploring as cell therapy for patients with RDEB. In terms of optimizing MSCs for clinical use, preconditioning of MSCs with TGF- β , TNF- α , and SDF-1 α , induces a simultaneous upregulation in *COL7A1*, *TSG-6*, and *CXCR4* which results in a six to eight-fold increase in *COL7A1* expression by MSCs (97). This pre-conditioning increased C7 levels towards the 30% of the amount of wild-type C7 believed to ameliorate the blistering seen in RDEB (75). Such preconditioning effects, however, have yet to be assessed therapeutically in humans.

8.3. Bone marrow transplantation

Following the effectiveness of bone marrow (BM) stem cells in murine RDEB (98,99), a clinical trial of whole bone marrow transplantation (BMT) was performed in children with RDEB.

In 2010, Wagner *et al.* (100) reported use of high dose chemotherapy to immunoablate individuals with RDEB to permit more reliable lymphohaematopoietic engraftment, followed by unfiltered whole bone marrow transplantation, usually from a tissue-matched sibling donor. Seven patients entered the trial and 6 underwent BMT. One patient died before the BMT because of heart failure, possibly related to cyclophosphamide toxicity and pre-existing renal failure. All RDEB subjects had more than 50% body surface area coverage with blisters and erosions. Following BMT, 3 subjects showed clinical improvement with only 10% BSA involvement and 3 showed an improvement with 25% BSA involvement. A further patient died 6 months post-transplant from infection secondary to graft failure. Of note, donor cells homed to injured skin with increased C7 expression seen at the DEJ in 5 of the 6 subjects. The subject that did not show evidence of increased C7 expression post-BMT was still reported to show an improvement in their clinical status, similar to that seen in the other 5 subjects that did show an increase in C7 expression. Clinical response seems to have been sustained; none of the treated subjects has been cured of their RDEB but

several have had markedly fewer blisters in follow up to 8 years post-BMT. Donor-skin chimerism was seen in the skin of BMT recipients (101). A substantial number of cells of donor origin were found in BMT recipient skin, confirming that donor cells home to injured skin in patients with severe RDEB. Donor cells of both haematopoietic (CD45+), and non-haematopoietic, non-endothelial cells (CD45-, CD31-) were found in the epidermis and dermis of BMT recipients, although donor non-haematopoietic cells were considered to be the most likely source of new C7 (101). Despite the increase in C7 expression, there was a lack of mature anchoring fibrils on transmission electron microscopy (TEM), although later evaluation will be needed given the several years anchoring fibril maturation may take.

Regarding the interconnectivity between BM cells and skin repair, the release of HMGB-1 from hypoxic keratinocytes and the mobilisation of Lin-/PDGFR α + epithelial progenitor cells from bone marrow to the circulation and differentiating into keratinocytes capable of generating new C7 in the skin, supports the potential mechanism of action of BMT (78). However, the homing of these cells to injured skin post-BMT has not yet been fully established. Reports suggest that the C-X-C type chemokine ligand 12 (CXCL12), known as stromal cell-derived factor 1 α (SDF-1 α), and its receptor, CXCR4 may direct the migration of progenitor cells to various tissues (102). The transcription factor hypoxia inducible factor-1 alpha, HIF-1 α , in endothelial cells in ischaemic tissue regulates the expression of SDF-1 α , enabling CXCR4+ progenitor cells to home from the circulation to target ischaemic tissue (103). Overall, despite the clinical data, the precise mechanism by which BMT leads to clinical improvement has not yet been fully elucidated. Of clinical significance, however, immunoablative conditioning in RDEB pre-BMT has been associated with mortality rates in excess of 25%. To lessen mortality, several refined stem cell transplantation protocols have been developed that focus on reduced intensity conditioning (RIC). Combination conditioning has been reduced from using busulfan, fludarabine, and cyclophosphamide to combination therapy with fludarabine and low doses of cyclophosphamide and radiation (101), although further refinements continue to be applied. Thus far, it appears that RIC is associated with less toxicity and relatively good disease amelioration, but published data are currently lacking.

8.4. Grafting revertant mosaicism skin/keratinocytes

In patients with various inherited cutaneous diseases, patches of spontaneously appearing normal skin can be seen where the inherited mutation has genetically corrected itself in those sites. This phenomenon is referred to as revertant mosaicism or "natural gene therapy" (104) and a key goal has been to try to exploit these natural events in the treatment of

RDEB. Thus far, revertant mosaicism has not been explored therapeutically in RDEB although in some forms of junctional EB, grafting of cultured revertant keratinocytes (105) or punch grafting of revertant skin has been undertaken, with sustained improvement in recipient mutant skin sites being demonstrated for the latter (106).

The opportunity to expand keratinocytes derived from a patch of revertant mosaicism offers a personalised and patient specific form of therapy. As these cells have naturally corrected part of the deleterious mutation, there is no need for further genetic manipulation. Gostynski *et al.* (105) isolated revertant keratinocytes from an individual with generalised intermediate junctional EB and expanded these into epidermal sheets to graft on to areas of mutant skin lacking an epidermis. The surgical approach led to successful grafting although functional benefits were not apparent. Of note, despite cultured keratinocytes displaying 30% reversion, when grafted, less than 3% of keratinocytes remained reverted in the graft; the reasons for this relative loss of reverted cells is not known. More successful was punch graft transplantation of revertant skin in an individual with junctional EB that resulted in successful transfer of the donor cell genotype and phenotype with enhanced expression of laminin-332 and better skin integrity maintained for at least 18 months (106). Nevertheless a key challenge is to find methods for higher *in vitro* expansion of revertant keratinocytes as well as being able to more readily identify the revertant skin patches (107). One new approach has been to generate inducible pluripotent stem cells (iPSCs) from revertant keratinocytes (see gene therapy section below) (108,109), which potentially then offers copious functional cells that can be differentiated into multiple tissue lineages.

8.5. Gene therapy

Gene therapy strategies in RDEB aim to provide therapeutic benefit through manipulation of DNA or RNA. Typically, viral mediated *ex vivo* gene transfer approaches have been used whereby the patient's skin cells are cultured, transduced with a viral vector encoding the transgene expressing the wild-type protein and then these gene modified cells can then either be transplanted back *via* grafting of epithelial sheets or skin equivalents (epidermis/dermis), or by intradermal injections (*e.g.* of genetically supplemented fibroblasts). Viral mediated gene transfer has been the preferred gene delivery method, firstly, due to the ability to deliver a transgene and integrate it into the host genomic DNA, and secondly because viral vector approaches achieve higher transduction efficiencies for longer-term gene expression. Gamma retroviral (RV) and lentiviral (LV) vectors have been the main delivery methods for RDEB gene therapy studies, despite the large size of the *COL7A1* cDNA (> 9 kb) (110-113). Regarding specific pre-clinical work for

RDEB, one study used an LV-mediated system to make intradermal injections of corrected patient-derived RDEB fibroblasts to restore C7 at the dermal-epidermal junction for 4 months in an RDEB skin model (111). Moreover, it was subsequently shown that direct intradermal injections of an LV vector containing *COL7A1* cDNA could produce stable expression of human C7 in fibroblasts and endothelial cells for at least 3 months in a murine model (114). To compensate for the large size of *COL7A1*, an RV vector with a truncated *COL7A1* "mini-gene" was first assessed (115). Immortalised RDEB keratinocytes could be transduced to express a mini-C7 protein product that improved cell motility, adhesion, and proliferation, although mini-gene therapy approaches have not been pursued to clinical trials.

The first clinical study of *ex vivo* gene therapy for EB was in an individual with junctional EB, with restoration of laminin-332 expression following RV-mediated transfection of epidermal stem cells with the *LAMB3* gene, leading to phenotypic correction in the grafted skin (116). Of note, follow up for more than 8 years has shown sustained synthesis of laminin-332 protein with no evidence of blistering, inflammation, tumourigenesis or immune response in the grafted area (117). In a second case, the same RV gene therapy protocol was used in an Austrian junctional EB patient in whom *ex vivo* skin gene therapy targeting autologous epidermal stem cells was used to produce five skin sheets each measuring 5 × 7 cm that were grafted onto wounded areas on the patient's thighs; clinical responses in this patient are still being evaluated (118).

The first gene therapy trial in RDEB involved grafting of *ex vivo* autologous *COL7A1* gene supplemented epidermal sheets in 4 adults in a phase I clinical trial. In this study, autologous keratinocytes were transduced with GMP grade gamma-RV containing full-length *COL7A1*. Autologous epidermal sheets measuring ~35cm² (approximately the size of a playing card) were grafted onto 6 wounds in each of the patients. No serious adverse events were reported and there was C7 expression at the dermal-epidermal junction on graft sites in 90% of biopsies at 3 months, 66% of biopsies at 6 months and 42% at 12 months. Wound healing was variable and generally waned over one year. Longer term follow-up will be required to ascertain long-term efficacy and safety (119).

The risk of insertional mutagenesis arising from use of certain classical viral vectors has led to a new generation of self-inactivating (SIN) viral vectors which incorporate deletion of the U3 region of the 3'-long terminal repeat that renders them unable to activate cellular genes in the host's genome. A SIN-LV-based vector was used to deliver full-length *COL7A1* cDNA sequence into patient-derived RDEB keratinocytes and fibroblasts (110). This approach gave close to 95% transduction efficiency and demonstrated persistent synthesis and secretion of normal C7 over a 5 month

observation period *in vitro* (110). These corrected cells were also able to produce normal anchoring fibrils when grafted onto immunodeficient mice. Other investigators are currently carrying out a clinical trial of SIN-LV vector *COL7A1* addition to autologous fibroblasts for intradermal injection (ClinicalTrials.gov identifier: NCT02493816), and others are developing a SIN-RV vector containing full length *COL7A1* with the aim being to transplant bioengineered skin containing genetically supplemented keratinocytes and fibroblasts (www.genegraft.eu).

As an alternative to viral-mediated transduction, a phage-mediated platform has been used to deliver *COL7A1* cDNA into patient-derived RDEB primary epidermal progenitor cells (120). The authors used a phiC31 phage integrase, which can integrate large (up to 10 kb) DNA sequences. The experimental data revealed relatively lower transfection efficiency rates (~ 45% at 2 days) compared to viral transduction methods, but through culture expansion and selection of C7-producing cells, a ~ 99% success rate after a 10-day selection period was noted. Moreover, C7 production by epidermal progenitor cells was suggested by persistent expression for 14 weeks, *i.e.* spanning multiple turnover cycles of keratinocytes. The same phiC31 phage integrase platform was subsequently used to correct patient-derived RDEB fibroblasts. Corrected fibroblasts were then injected into an RDEB skin model and were shown to restore C7 expression in the skin (121). Nevertheless, the requirement to include the phiC31 integrase gene, the lack of responsiveness to endogenous gene regulation, and the potential for random insertional mutagenesis may be limiting factors for phage therapy.

Cationic polymers such as linear poly (β -amino ester)s (LPAEs) have also emerged as an effective gene delivery vector. Branched poly (β -amino ester)s (HPAEs) have a three-dimensional spatial structure and are thought to improve the interaction of polymers with DNA, prevent DNA degradation by enzymes and increase cellular uptake of polyplexes. HPAEs have not been developed for gene delivery as yet, as synthesising these highly branched polymers remains a technical challenge. A novel design of the HPAEs has been derived from the functional LPAE components to see whether this may provide an effective gene delivery vector. This has been assessed *in vivo* in various cell types including RDEB keratinocytes to deliver therapeutic *COL7A1* cDNA (122).

Gene silencing technologies such as RNA interference (RNAi) are useful in dominant forms of DEB, if designed to knockdown the mutant allele without silencing the wild-type allele, with pre-clinical data to support therapeutic use of such an approach (123,124). Another methodology, pertinent mainly to RDEB but possibly also dominant disease, is to try to modulate splicing of pre-messenger RNA to induce

skipping of the mutated exon. Using 2'-O-methyl antisense oligoribonucleotides (AONs) in an RDEB skin equivalent xenograft model, one or two subcutaneous injections of AONs at doses ranging from 400 μ g up to 1 mg was able to induce skipping of exons containing loss-of-function mutations (in exons 73 and 80) and thereby restore C7 expression and anchoring fibril formation (125). A further method is to apply spliceosome-mediated RNA trans-splicing (SMaRT) to address target mutations at a post transcriptional level. Splicing is induced *in trans* between the exogenous RNA and target endogenous pre-mRNA *via* an engineered RNA trans-splicing molecule (RTM). Specifically, RV transduction of RDEB keratinocytes with a 3' pre-trans-splicing molecule resulted in correction of full-length C7 expression (126). Transduced cells showed normal localisation of C7 at the basement membrane zone in skin equivalents with assembly into anchoring fibril-like structures, *i.e.* demonstrating correction of an RDEB phenotype *in vitro* (126). In further work, a 5' RTM capable of replacing *COL7A1* exons 1 to 15 in murine keratinocytes was injected into the skin of wild-type mice using a gene gun with vector delivery and expression in the skin (127).

Approximately 15% of all pathogenic mutations in *COL7A1* involve premature termination codons (PTCs) that lead to truncated proteins and/or nonsense-mediated mRNA decay (128). Both *in vitro* and *in vivo* studies have revealed that aminoglycoside antibiotics can suppress primary PTCs and produce some degree of full length functional protein in genetic disorders such as cystic fibrosis (CF) and Duchenne's muscular dystrophy (DMD) (129,130). In RDEB, preclinical analysis has been performed using two RDEB keratinocyte cell lines harbouring nonsense mutations and primary fibroblast cultures from two RDEB patients with nonsense mutations. Aminoglycosides (G418, gentamicin, and paramomycin) were able to induce PTC read-through and restore functional full-length C7. Aminoglycoside therapy may provide a non-invasive option in treating RDEB patients that carry nonsense mutations but has not yet been trialled. Potential toxicity and the extent of the readthrough necessary to generate functional correction, however, remain important considerations that may limit immediate clinical translation.

Genomic editing techniques including zinc-finger nucleases (ZFNs), meganucleases (MN), transcription activator-like effector nucleases (TALENs) (131,132) and the clustered regularly interspaced short palindromic repeats (CRISPR)-Cas9 nuclease system are being developed (133), some or all of which may have relevance to RDEB therapeutics.

Moreover, the advent reprogramming of somatic cells into induced pluripotent stem cells (iPSCs) that can differentiate into any cell type, is an exciting new development in RDEB therapy (134). It is possible to correct RDEB fibroblasts through homologous recombination using transcription activator-like effector

nucleases (TALENs) and then reprogram these into iPSCs, which then differentiate into keratinocytes (135). Murine studies have also successfully generated iPSCs in culture from multipotent keratinocyte lineages capable of forming a fully developed epidermis (136). Subsequently, others have reported successful generation of iPSCs from healthy human skin fibroblasts and individuals with RDEB (137). Another study took a different approach using direct injections and teratoma formation which allows spontaneous differentiation of iPSCs into an epidermis (138). Regarding new therapeutic opportunities, an approach in which iPSCs generated from naturally corrected revertant RDEB cells could be used to enable the production of autologous epithelial and mesenchymal cells, perhaps paving the way for personalised therapy in EB (108,109).

8.6. Protein therapy

Given that the essential skin pathology in RDEB is a lack of C7 in epidermal basement membrane, C7 protein replacement therapy has been evaluated using animal models for preclinical studies. Initial studies successfully demonstrated that intradermal injections of recombinant human C7 can lead to incorporation of the new protein specifically into basement membrane of *Col7a1* null mice, resulting in an improvement in the blistering phenotype for up to 2 months (139). Furthermore, topical application of human recombinant C7 accelerated wound healing in mice (140), and intravenously administered rC7 homed to engrafted RDEB mouse skin and restored C7, anchoring fibrils, and epidermal-dermal adherence (139,141).

Concerning larger animal studies, intravenous administration of C7 in a spontaneous animal model of inbred mini Retriever dogs with mild RDEB revealed no side effects and led to reduced wound erythema and blistering (142). Initially, no serious immunological reactions were observed, and although anti-C7 antibodies were detectable in serum, none was shown to bind to the skin or exacerbate blistering (143,144). The development of human C7 protein trials was expected thereafter, although thus far additional possible toxicology concerns have stalled clinical application, and further research will be required to assess the efficacy and safety of this therapy before clinical testing in patients with RDEB.

9. The Future

There is an urgent need for curative therapies for genetic disorders like RDEB that carry significant morbidity and mortality. In future, optimal treatment of RDEB will most likely involve combinations of drug, small molecule, gene, cell and protein therapies, with the collective ambition of reducing disease burden and compensating for, or repairing, the inherent skin pathology underscoring the blistering.

References

1. Fine JD, Bruckner-Tuderman L, Eady RA, *et al.* Inherited epidermolysis bullosa: Updated recommendations on diagnosis and classification. *J Am Acad Dermatol.* 2014; 70:1103-1126.
2. Hsu CK, Wang SP, Lee JY, McGrath JA. Treatment of hereditary epidermolysis bullosa: Updates and future prospects. *Am J Clin Dermatol.* 2014; 15:1-6.
3. Sakai LY, Keene DR, Morris NP, Burgeson RE. Type VII collagen is a major structural component of anchoring fibrils. *J Cell Biol.* 1986; 103:1577-1586.
4. Shimizu H, McGrath JA, Christiano AM, Nishikawa T, Uitto J. Molecular basis of recessive dystrophic epidermolysis bullosa: Genotype/phenotype correlation in a case of moderate clinical severity. *J Invest Dermatol.* 1996; 106:119-124.
5. Dang N, Murrell DF. Mutation analysis and characterization of *COL7A1* mutations in dystrophic epidermolysis bullosa. *Exp Dermatol.* 2008; 17:553-568.
6. Leigh IM, Eady RA, Heagerty AH, Purkis PE, Whitehead PA, Burgeson RE. Type VII collagen is a normal component of epidermal basement membrane, which shows altered expression in recessive dystrophic epidermolysis bullosa. *J Invest Dermatol.* 1988; 90:639-642.
7. Denyer J. Managing pain in children with epidermolysis bullosa. *Nurs Times.* 2012; 108:21-23.
8. Fine JD, Mellerio JE. Extracutaneous manifestations and complications of inherited epidermolysis bullosa: Part II. Other organs. *J Am Acad Dermatol.* 2009; 61:387-402; quiz 403-404.
9. Goldschneider KR, Good J, Harrop E, Lioffi C, Lynch-Jordan A, Martinez AE, Maxwell LG, Stanko-Lopp D, Dystrophic Epidermolysis Bullosa Research Association I. Pain care for patients with epidermolysis bullosa: Best care practice guidelines. *BMC Med.* 2014; 12:178.
10. Freeman EB, Kogelmeier J, Martinez AE, Mellerio JE, Haynes L, Sebire NJ, Lindley KJ, Shah N. Gastrointestinal complications of epidermolysis bullosa in children. *Br J Dermatol.* 2008; 158:1308-1314.
11. Chiu YK, Prendiville JS, Bennett SM, Montgomery CJ, Oberlander TF. Pain management of junctional epidermolysis bullosa in an 11-year-old boy. *Pediatr Dermatol.* 1999; 16:465-468.
12. van Scheppingen C, Lettinga AT, Duipmans JC, Maathuis CG, Jonkman MF. Main problems experienced by children with epidermolysis bullosa: A qualitative study with semi-structured interviews. *Acta Derm Venereol.* 2008; 88:143-150.
13. Daniel C, Adeduntan R, Gorell ES, Lucky AW, Paller AS, Bruckner A, Pope E, Morel KD, Levy ML, Li S, Gilmore ES, Lane AT. Prevalence and characterization of pruritus in epidermolysis bullosa. *Pediatr Dermatol.* 2015; 32:53-59.
14. Wright JT, Fine JD, Johnson L. Dental caries risk in hereditary epidermolysis bullosa. *Pediatr Dent.* 1994; 16:427-432.
15. Fortuna G, Aria M, Cepeda-Valdes R, Pollio A, Moreno-Trevino MG, Salas-Alanis JC. Clinical features of gingival lesions in patients with dystrophic epidermolysis bullosa: A cross-sectional study. *Aust Dent J.* 2015; 60:18-23.
16. Crawford EG, Jr, Burkes EJ, Jr, Briggaman RA. Hereditary epidermolysis bullosa: Oral manifestations and dental therapy. *Oral Surg Oral Med Oral Pathol.* 1976;

- 42:490-500.
17. Wright JT. Comprehensive dental care and general anesthetic management of hereditary epidermolysis bullosa. A review of fourteen cases. *Oral Surg Oral Med Oral Pathol.* 1990; 70:573-578.
 18. Marini I, Vecchiet F. Sucralfate: A help during oral management in patients with epidermolysis bullosa. *J Periodontol.* 2001; 72:691-695.
 19. Colomb V, Bourdon-Lannoy E, Lambe C, Sauvat F, Hadj Rabia S, Teillac D, De Prost Y, Bodemer C. Nutritional outcome in children with severe generalized recessive dystrophic epidermolysis bullosa: A short- and long-term evaluation of gastrostomy and enteral feeding. *Br J Dermatol.* 2012; 166:354-361.
 20. Fewtrell MS, Allgrove J, Gordon I, Brain C, Atherton D, Harper J, Mellerio JE, Martinez AE. Bone mineralization in children with epidermolysis bullosa. *Br J Dermatol.* 2006; 154:959-962.
 21. Martinez AE, Mellerio JE. Osteopenia and osteoporosis in epidermolysis bullosa. *Dermatol Clin.* 2010; 28:353-355.
 22. Martinez AE, Allgrove J, Brain C. Growth and pubertal delay in patients with epidermolysis bullosa. *Dermatol Clin.* 2010; 28:357-359.
 23. Pillay E. Epidermolysis bullosa. Part 1: Causes, presentation and complications. *Br J Nurs.* 2008; 17:292-296.
 24. Pope E, Lara-Corrales I, Mellerio J, Martinez A, Schultz G, Burrell R, Goodman L, Coutts P, Wagner J, Allen U, Sibbald G. A consensus approach to wound care in epidermolysis bullosa. *J Am Acad Dermatol.* 2012; 67:904-917.
 25. Marin-Bertolin S, Amaya Valero JV, Neira Gimenez C, Marquina Vila P, Amorrortu-Velayos J. Surgical management of hand contractures and pseudosyndactyly in dystrophic epidermolysis bullosa. *Ann Plast Surg.* 1999; 43:555-559.
 26. Bernardis C, Box R. Surgery of the hand in recessive dystrophic epidermolysis bullosa. *Dermatol Clin.* 2010; 28:335-341, xi.
 27. McGrath JA, O'Grady A, Mayou BJ, Eady RA. Mitten deformity in severe generalized recessive dystrophic epidermolysis bullosa: Histological, immunofluorescence, and ultrastructural study. *J Cutan Pathol.* 1992; 19:385-389.
 28. Terrill PJ, Mayou BJ, Pemberton J. Experience in the surgical management of the hand in dystrophic epidermolysis bullosa. *Br J Plast Surg.* 1992; 45:435-442.
 29. Fine JD, Johnson LB, Weiner M, Li KP, Suchindran C. Epidermolysis bullosa and the risk of life-threatening cancers: The National EB Registry experience, 1986-2006. *J Am Acad Dermatol.* 2009; 60:203-211.
 30. Tomita Y, Sato-Matsumura KC, Sawamura D, Matsumura T, Shimizu H. Simultaneous occurrence of three squamous cell carcinomas in a recessive dystrophic epidermolysis bullosa patient. *Acta Derm Venereol.* 2003; 83:225-226.
 31. Mellerio JE, Robertson SJ, Bernardis C, *et al.* Management of cutaneous squamous cell carcinoma in patients with epidermolysis bullosa: Best clinical practice guidelines. *Br J Dermatol.* 2016; 174:56-67.
 32. Lentz SR, Raish RJ, Orlowski EP, Marion JM. Squamous cell carcinoma in epidermolysis bullosa. Treatment with systemic chemotherapy. *Cancer.* 1990; 66:1276-1278.
 33. Wechsler HL, Krugh FJ, Domonkos AN, Scheen SR, Davidson CL, Jr. Polydysplastic epidermolysis bullosa and development of epidermal neoplasms. *Arch Dermatol.* 1970; 102:374-380.
 34. Mallipeddi R. Epidermolysis bullosa and cancer. *Clin Exp Dermatol.* 2002; 27:616-623.
 35. Schwartz RA, Birnkrant AP, Rubenstein DJ, Kim U, Burgess GH, Stoll HL, Jr, Chai SW, Southwick GJ, Milgrom H. Squamous cell carcinoma in dominant type epidermolysis bullosa dystrophica. *Cancer.* 1981; 47:615-620.
 36. Gebbia V, Giuliani F, Valori VM, Agueli R, Colucci G, Maiello E. Cetuximab in squamous cell head and neck carcinomas. *Ann Oncol.* 2007; 18 Suppl 6:vi5-7.
 37. Sharafinski ME, Ferris RL, Ferrone S, Grandis JR. Epidermal growth factor receptor targeted therapy of squamous cell carcinoma of the head and neck. *Head Neck.* 2010; 32:1412-1421.
 38. Maubec E, Petrow P, Scheer-Senyarich I, *et al.* Phase II study of cetuximab as first-line single-drug therapy in patients with unresectable squamous cell carcinoma of the skin. *J Clin Oncol.* 2011; 29:3419-3426.
 39. Kim M, Li M, Intong LR, Tran K, Melbourne W, Marucci D, Bucci J, de Souza P, Mallesara G, Murrell DF. Use of cetuximab as an adjuvant agent to radiotherapy and surgery in recessive dystrophic epidermolysis bullosa with squamous cell carcinoma. *Br J Dermatol.* 2013; 169:208-210.
 40. Arnold AW, Bruckner-Tuderman L, Zuger C, Itin PH. Cetuximab therapy of metastasizing cutaneous squamous cell carcinoma in a patient with severe recessive dystrophic epidermolysis bullosa. *Dermatology.* 2009; 219:80-83.
 41. Fine JD, Johnson LB, Weiner M, Stein A, Suchindran C. Chemoprevention of squamous cell carcinoma in recessive dystrophic epidermolysis bullosa: Results of a phase 1 trial of systemic isotretinoin. *J Am Acad Dermatol.* 2004; 50:563-571.
 42. Shinkuma S, Sawamura D, Fujita Y, Kawasaki H, Nakamura H, Inoie M, Nishie W, Shimizu H. Long-term follow-up of cultured epidermal autograft in a patient with recessive dystrophic epidermolysis bullosa. *Acta Derm Venereol.* 2014; 94:98-99.
 43. Fivenson DP, Scherschun L, Cohen LV. Apligraf in the treatment of severe mitten deformity associated with recessive dystrophic epidermolysis bullosa. *Plast Reconstr Surg.* 2003; 112:584-588.
 44. Falabella AF, Valencia IC, Eaglstein WH, Schachner LA. Tissue-engineered skin (Apligraf) in the healing of patients with epidermolysis bullosa wounds. *Arch Dermatol.* 2000; 136:1225-1230.
 45. Hasegawa T, Suga Y, Mizoguchi M, Ikeda S, Ogawa H, Kubo K, Matsui H, Kagawa S, Kuroyanagi Y. Clinical trial of allogeneic cultured dermal substitute for the treatment of intractable skin ulcers in 3 patients with recessive dystrophic epidermolysis bullosa. *J Am Acad Dermatol.* 2004; 50:803-804.
 46. Natsuga K, Sawamura D, Goto M, Homma E, Goto-Ohguchi Y, Aoyagi S, Akiyama M, Kuroyanagi Y, Shimizu H. Response of intractable skin ulcers in recessive dystrophic epidermolysis bullosa patients to an allogeneic cultured dermal substitute. *Acta Derm Venereol.* 2010; 90:165-169.
 47. Falabella AF, Schachner LA, Valencia IC, Eaglstein WH. The use of tissue-engineered skin (Apligraf) to treat a newborn with epidermolysis bullosa. *Arch Dermatol.* 1999; 135:1219-1222.
 48. Niknejad H, Peirovi H, Jorjani M, Ahmadiani A, Ghanavi

- J, Seifalian AM. Properties of the amniotic membrane for potential use in tissue engineering. *Eur Cell Mater.* 2008; 15:88-99.
49. Lo V, Lara-Corrales I, Stuparich A, Pope E. Amniotic membrane grafting in patients with epidermolysis bullosa with chronic wounds. *J Am Acad Dermatol.* 2010; 62:1038-1044.
 50. Martinez Pardo ME, Reyes Frias ML, Ramos Duron LE, Gutierrez Salgado E, Gomez JC, Marin MA, Luna Zaragoza D. Clinical application of amniotic membranes on a patient with epidermolysis bullosa. *Ann Transplant.* 1999; 4:68-73.
 51. Gibbons GW. Grafix[®], a cryopreserved placental membrane, for the treatment of chronic/stalled wounds. *Adv Wound Care (New Rochelle).* 2015; 4:534-544.
 52. Nevala-Plagemann C, Lee C, Tolar J. Placenta-based therapies for the treatment of epidermolysis bullosa. *Cytotherapy.* 2015; 17:786-795.
 53. Lavery LA, Fulmer J, Shebetka KA, Regulski M, Vayser D, Fried D, Kashefsky H, Owings TM, Nadarajah J; Grafix Diabetic Foot Ulcer Study Group. The efficacy and safety of Grafix[®] for the treatment of chronic diabetic foot ulcers: Results of a multi-centre, controlled, randomised, blinded, clinical trial. *Int Wound J.* 2014; 11:554-560.
 54. Dagregorio G, Guillet G. Artificial skin as a valuable adjunct to surgical treatment of a large squamous cell carcinoma in a patient with epidermolysis bullosa. *Dermatol Surg.* 2005; 31:474-476.
 55. Wiegand C, Schonfelder U, Abel M, Ruth P, Kaatz M, Hipler UC. Protease and pro-inflammatory cytokine concentrations are elevated in chronic compared to acute wounds and can be modulated by collagen type I *in vitro*. *Arch Dermatol Res.* 2010; 302:419-428.
 56. Gorell ES, Leung TH, Khuu P, Lane AT. Purified type I collagen wound matrix improves chronic wound healing in patients with recessive dystrophic epidermolysis bullosa. *Pediatr Dermatol.* 2015; 32:220-225.
 57. Fine JD. Epidermolysis bullosa: A genetic disease of altered cell adhesion and wound healing, and the possible clinical utility of topically applied thymosin beta4. *Ann N Y Acad Sci.* 2007; 1112:396-406.
 58. Margolis DJ, Morris LM, Papadopoulos M, Weinberg L, Filip JC, Lang SA, Vaikunth SS, Crombleholme TM. Phase I study of H5.020CMV.PDGF-beta to treat venous leg ulcer disease. *Mol Ther.* 2009; 17:1822-1829.
 59. Ma C, Hernandez MA, Kirkpatrick VE, Liang LJ, Nouvong AL, Gordon, II. Topical platelet-derived growth factor vs placebo therapy of diabetic foot ulcers offloaded with windowed casts: A randomized, controlled trial. *Wounds.* 2015; 27:83-91.
 60. Pearson RW. Studies on the pathogenesis of epidermolysis bullosa. *J Invest Dermatol.* 1962; 39:551-575.
 61. Bauer EA, Cooper TW, Tucker DR, Esterly NB. Phenytoin therapy of recessive dystrophic epidermolysis bullosa. Clinical trial and proposed mechanism of action on collagenase. *N Engl J Med.* 1980; 303:776-781.
 62. Caldwell-Brown D, Stern RS, Lin AN, Carter DM. Lack of efficacy of phenytoin in recessive dystrophic epidermolysis bullosa. *Epidermolysis Bullosa Study Group.* *N Engl J Med.* 1992; 327:163-167.
 63. White JE. Minocycline for dystrophic epidermolysis bullosa. *Lancet.* 1989; 1:966.
 64. Golub LM, Wolff M, Lee HM, McNamara TF, Ramamurthy NS, Zambon J, Ciancio S. Further evidence that tetracyclines inhibit collagenase activity in human crevicular fluid and from other mammalian sources. *J Periodontal Res.* 1985; 20:12-23.
 65. Lettner T, Lang R, Bauer JW, Wally V. Increased levels of matrix metalloproteinase-9 and interleukin-8 in blister fluids of dystrophic and junctional epidermolysis bullosa patients. *J Eur Acad Dermatol Venereol.* 2015; 29:396-398.
 66. Leung J, Kuzel P, Kurian A, Brassard A. A Case of Dominant Dystrophic Epidermolysis Bullosa Responding Well to an Old Medication. *JAMA Dermatol.* 2015; 151:1264-1265.
 67. Esterly NB, Furey NL, Flanagan LE. The effect of antimicrobial agents on leukocyte chemotaxis. *J Invest Dermatol.* 1978; 70:51-55.
 68. Lara-Corrales I, Parkin PC, Stephens D, Hamilton J, Koren G, Weinstein M, Sibbald RG, Pope E. The efficacy of trimethoprim in wound healing of patients with epidermolysis bullosa: A feasibility trial. *J Am Acad Dermatol.* 2012; 66:264-270.
 69. Changotade SI, Assoumou A, Gueniche F, Fioretti F, Segulier S, de Prost Y, Bodemer C, Godeau G, Senni K. Epigallocatechin gallate's protective effect against MMP7 in recessive dystrophic epidermolysis bullosa patients. *J Invest Dermatol.* 2007; 127:821-828.
 70. Chiaverini C, Roger C, Fontas E, Bourrat E, Bourdon-Lanoy E, Labreze C, Mazereeuw J, Vabres P, Bodemer C, Lacour JP. Oral epigallocatechin-3-gallate for treatment of dystrophic epidermolysis bullosa: A multicentre, randomized, crossover, double-blind, placebo-controlled clinical trial. *Orphanet J Rare Dis.* 2016; 11:31.
 71. del-Rio E. Prevention of blisters in dystrophic recessive epidermolysis bullosa with cyclosporine. *J Am Acad Dermatol.* 1993; 29:1038-1039.
 72. El-Darouti MA, Fawzy MM, Amin IM, Abdel Hay RM, Hegazy RA, Abdel Halim DM. Mycophenolate mofetil: A novel immunosuppressant in the treatment of dystrophic epidermolysis bullosa, a randomized controlled trial. *J Dermatolog Treat.* 2013; 24:422-426.
 73. Gubinelli E, Angelo C, Pacifico V. A case of dystrophic epidermolysis bullosa improved with etanercept for concomitant psoriatic arthritis. *Am J Clin Dermatol.* 2010; 11 Suppl 1:53-54.
 74. Fine JD, Manes B, Frangoul H. Systemic granulocyte colony-stimulating factor (G-CSF) enhances wound healing in dystrophic epidermolysis bullosa (DEB): Results of a pilot trial. *J Am Acad Dermatol.* 2015; 73:56-61.
 75. Fritsch A, Loeckermann S, Kern JS, Braun A, Bosl MR, Bley TA, Schumann H, von Elverfeldt D, Paul D, Erlacher M, Berens von Rautenfeld D, Hausser I, Fassler R, Bruckner-Tuderman L. A hypomorphic mouse model of dystrophic epidermolysis bullosa reveals mechanisms of disease and response to fibroblast therapy. *J Clin Invest.* 2008; 118:1669-1679.
 76. Nystrom A, Thriene K, Mittapalli V, Kern JS, Kiritsi D, Dengjel J, Bruckner-Tuderman L. Losartan ameliorates dystrophic epidermolysis bullosa and uncovers new disease mechanisms. *EMBO Mol Med.* 2015; 7:1211-1228.
 77. Odorisio T, Di Salvio M, Orecchia A, Di Zenzo G, Piccinni E, Cianfarani F, Travaglione A, Uva P, Bellei B, Conti A, Zambruno G, Castiglia D. Monozygotic twins discordant for recessive dystrophic epidermolysis bullosa phenotype highlight the role of TGF-beta signalling in modifying disease severity. *Hum Mol Genet.* 2014;

- 23:3907-3922.
78. Tamai K, Yamazaki T, Chino T, *et al.* PDGFR α -positive cells in bone marrow are mobilized by high mobility group box 1 (HMGB1) to regenerate injured epithelia. *Proc Natl Acad Sci U S A.* 2011; 108:6609-6614.
 79. Stanley JR, Rubinstein N, Klaus-Kovtun V. Epidermolysis bullosa acquisita antigen is synthesized by both human keratinocytes and human dermal fibroblasts. *J Invest Dermatol.* 1985; 85:542-545.
 80. Wong T, Gammon L, Liu L, Mellerio JE, Dopping-Hepenstal PJ, Pacy J, Elia G, Jeffery R, Leigh IM, Navsaria H, McGrath JA. Potential of fibroblast cell therapy for recessive dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2008; 128:2179-2189.
 81. Kern JS, Loeckermann S, Fritsch A, Hausser I, Roth W, Magin TM, Mack C, Muller ML, Paul O, Ruther P, Bruckner-Tuderman L. Mechanisms of fibroblast cell therapy for dystrophic epidermolysis bullosa: High stability of collagen VII favors long-term skin integrity. *Mol Ther.* 2009; 17:1605-1615.
 82. Nagy N, Almaani N, Tanaka A, Lai-Cheong JE, Techanukul T, Mellerio JE, McGrath JA. HB-EGF induces COL7A1 expression in keratinocytes and fibroblasts: Possible mechanism underlying allogeneic fibroblast therapy in recessive dystrophic epidermolysis Bullosa. *J Invest Dermatol.* 2011; 131:1771-1774.
 83. Venugopal SS, Yan W, Frew JW, Cohn HI, Rhodes LM, Tran K, Melbourne W, Nelson JA, Sturm M, Fogarty J, Marinkovich MP, Igawa S, Ishida-Yamamoto A, Murrell DF. A phase II randomized vehicle-controlled trial of intradermal allogeneic fibroblasts for recessive dystrophic epidermolysis bullosa. *J Am Acad Dermatol.* 2013; 69:898-908.
 84. Petrof G, Martinez-Queipo M, Mellerio JE, Kemp P, McGrath JA. Fibroblast cell therapy enhances initial healing in recessive dystrophic epidermolysis bullosa wounds: Results of a randomized, vehicle-controlled trial. *Br J Dermatol.* 2013; 169:1025-1033.
 85. Najar M, Raicevic G, Boufker HI, Fayyad Kazan H, De Bruyn C, Meuleman N, Bron D, Toungouz M, Lagneaux L. Mesenchymal stromal cells use PGE2 to modulate activation and proliferation of lymphocyte subsets: Combined comparison of adipose tissue, Wharton's Jelly and bone marrow sources. *Cell Immunol.* 2010; 264:171-179.
 86. Montesinos JJ, Flores-Figueroa E, Castillo-Medina S, Flores-Guzman P, Hernandez-Estevez E, Fajardo-Orduna G, Orozco S, Mayani H. Human mesenchymal stromal cells from adult and neonatal sources: Comparative analysis of their morphology, immunophenotype, differentiation patterns and neural protein expression. *Cytotherapy.* 2009; 11:163-176.
 87. Conget P, Rodriguez F, Kramer S, Allers C, Simon V, Palisson F, Gonzalez S, Yubero MJ. Replenishment of type VII collagen and re-epithelialization of chronically ulcerated skin after intradermal administration of allogeneic mesenchymal stromal cells in two patients with recessive dystrophic epidermolysis bullosa. *Cytotherapy.* 2010; 12:429-431.
 88. El-Darouti M, Fawzy M, Amin I, Abdel Hay R, Hegazy R, Gabr H, El Maadawi Z. Treatment of dystrophic epidermolysis bullosa with bone marrow non-hematopoietic stem cells: A randomized controlled trial. *Dermatol Ther.* 2016; 29:96-100.
 89. Petrof G, Lwin SM, Martinez-Queipo M, Abdul-Wahab A, Tso S, Mellerio JE, Slaper-Cortenbach I, Boelens JJ, Tolar J, Veys P, Ofuya M, Peacock JL, Martinez AE, McGrath JA. Potential of Systemic Allogeneic Mesenchymal Stromal Cell Therapy for Children with Recessive Dystrophic Epidermolysis Bullosa. *J Invest Dermatol.* 2015; 135:2319-2321.
 90. Kuhl T, Mezger M, Hausser I, Handgretinger R, Bruckner-Tuderman L, Nystrom A. High local concentrations of intradermal MSCs restore skin integrity and facilitate wound healing in dystrophic epidermolysis bullosa. *Mol Ther.* 2015; 23:1368-1379.
 91. Prockop DJ. Repair of tissues by adult stem/progenitor cells (MSCs): Controversies, myths, and changing paradigms. *Mol Ther.* 2009; 17:939-946.
 92. Qi Y, Jiang D, Sindrilaru A, *et al.* TSG-6 released from intradermally injected mesenchymal stem cells accelerates wound healing and reduces tissue fibrosis in murine full-thickness skin wounds. *J Invest Dermatol.* 2014; 134:526-537.
 93. Wang M, Windgassen D, Papoutsakis ET. Comparative analysis of transcriptional profiling of CD3+, CD4+ and CD8+ T cells identifies novel immune response players in T-cell activation. *BMC Genomics.* 2008; 9:225.
 94. Ren G, Zhang L, Zhao X, Xu G, Zhang Y, Roberts AI, Zhao RC, Shi Y. Mesenchymal stem cell-mediated immunosuppression occurs *via* concerted action of chemokines and nitric oxide. *Cell Stem Cell.* 2008; 2:141-150.
 95. Liao Y, Geyer MB, Yang AJ, Cairo MS. Cord blood transplantation and stem cell regenerative potential. *Exp Hematol.* 2011; 39:393-412.
 96. Liao Y, Itoh M, Yang A, Zhu H, Roberts S, Hight AM, Latshaw S, Mitchell K, van de Ven C, Christiano A, Cairo MS. Human cord blood-derived unrestricted somatic stem cells promote wound healing and have therapeutic potential for patients with recessive dystrophic epidermolysis bullosa. *Cell Transplant.* 2014; 23:303-317.
 97. Perdoni C, McGrath JA, Tolar J. Preconditioning of mesenchymal stem cells for improved transplantation efficacy in recessive dystrophic epidermolysis bullosa. *Stem Cell Res Ther.* 2014; 5:121.
 98. Tolar J, Ishida-Yamamoto A, Riddle M, McElmurry RT, Osborn M, Xia L, Lund T, Slattery C, Uitto J, Christiano AM, Wagner JE, Blazar BR. Amelioration of epidermolysis bullosa by transfer of wild-type bone marrow cells. *Blood.* 2009; 113:1167-1174.
 99. Chino T, Tamai K, Yamazaki T, Otsuru S, Kikuchi Y, Nimura K, Endo M, Nagai M, Uitto J, Kitajima Y, Kaneda Y. Bone marrow cell transfer into fetal circulation can ameliorate genetic skin diseases by providing fibroblasts to the skin and inducing immune tolerance. *Am J Pathol.* 2008; 173:803-814.
 100. Wagner JE, Ishida-Yamamoto A, McGrath JA, Hordinsky M, Keene DR, Woodley DT, Chen M, Riddle MJ, Osborn MJ, Lund T, Dolan M, Blazar BR, Tolar J. Bone marrow transplantation for recessive dystrophic epidermolysis bullosa. *N Engl J Med.* 2010; 363:629-639.
 101. Tolar J, Wagner JE. Allogeneic blood and bone marrow cells for the treatment of severe epidermolysis bullosa: Repair of the extracellular matrix. *Lancet.* 2013; 382:1214-1223.
 102. Iinuma S, Aikawa E, Tamai K, Fujita R, Kikuchi Y, Chino T, Kikuta J, McGrath JA, Uitto J, Ishii M, Iizuka H,

- Kaneda Y. Transplanted bone marrow-derived circulating PDGFRalpha+ cells restore type VII collagen in recessive dystrophic epidermolysis bullosa mouse skin graft. *J Immunol.* 2015; 194:1996-2003.
103. Ceradini DJ, Kulkarni AR, Callaghan MJ, Tepper OM, Bastidas N, Kleinman ME, Capla JM, Galiano RD, Levine JP, Gurtner GC. Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. *Nat Med.* 2004; 10:858-864.
 104. Jonkman MF, Scheffer H, Stulp R, Pas HH, Nijenhuis M, Heeres K, Owaribe K, Pulkkinen L, Uitto J. Revertant mosaicism in epidermolysis bullosa caused by mitotic gene conversion. *Cell.* 1997; 88:543-551.
 105. Gostynski A, Deviaene FC, Pasmooij AM, Pas HH, Jonkman MF. Adhesive stripping to remove epidermis in junctional epidermolysis bullosa for revertant cell therapy. *Br J Dermatol.* 2009; 161:444-447.
 106. Yuen WY, Huizinga J, Jonkman MF. Punch grafting of chronic ulcers in patients with laminin-332-deficient, non-Herlitz junctional epidermolysis bullosa. *J Am Acad Dermatol.* 2013; 68:93-97.e2.
 107. Pasmooij AM, Pas HH, Deviaene FC, Nijenhuis M, Jonkman MF. Multiple correcting *COL7A1* mutations in patients with revertant mosaicism of epidermolysis bullosa. *Am J Hum Genet.* 2005; 77:727-740.
 108. Tolar J, McGrath JA, Xia L, Riddle MJ, Lees CJ, Eide C, Keene DR, Liu L, Osborn MJ, Lund TC, Blazar BR, Wagner JE. Patient-specific naturally gene-reverted induced pluripotent stem cells in recessive dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2014; 134:1246-1254.
 109. Umegaki-Arao N, Pasmooij AM, Itoh M, Cerise JE, Guo Z, Levy B, Gostynski A, Rothman LR, Jonkman MF, Christiano AM. Induced pluripotent stem cells from human revertant keratinocytes for the treatment of epidermolysis bullosa. *Sci Transl Med.* 2014; 6:264ra164.
 110. Chen M, Kasahara N, Keene DR, Chan L, Hoeffler WK, Finlay D, Barcova M, Cannon PM, Mazurek C, Woodley DT. Restoration of type VII collagen expression and function in dystrophic epidermolysis bullosa. *Nat Genet.* 2002; 32:670-675.
 111. Woodley DT, Krueger GG, Jorgensen CM, Fairley JA, Atha T, Huang Y, Chan L, Keene DR, Chen M. Normal and gene-corrected dystrophic epidermolysis bullosa fibroblasts alone can produce type VII collagen at the basement membrane zone. *J Invest Dermatol.* 2003; 121:1021-1028.
 112. Baldeschi C, Gache Y, Rattenholl A, Bouille P, Danos O, Ortonne JP, Bruckner-Tuderman L, Meneguzzi G. Genetic correction of canine dystrophic epidermolysis bullosa mediated by retroviral vectors. *Hum Mol Genet.* 2003; 12:1897-1905.
 113. Titeux M, Pendaries V, Zanta-Boussif MA, Decha A, Pironon N, Tonasso L, Mejia JE, Brice A, Danos O, Hovnanian A. SIN retroviral vectors expressing COL7A1 under human promoters for *ex vivo* gene therapy of recessive dystrophic epidermolysis bullosa. *Mol Ther.* 2010; 18:1509-1518.
 114. Woodley DT, Keene DR, Atha T, Huang Y, Ram R, Kasahara N, Chen M. Intradermal injection of lentiviral vectors corrects regenerated human dystrophic epidermolysis bullosa skin tissue *in vivo*. *Mol Ther.* 2004; 10:318-326.
 115. Chen M, O'Toole EA, Muellenhoff M, Medina E, Kasahara N, Woodley DT. Development and characterization of a recombinant truncated type VII collagen "minigene". Implication for gene therapy of dystrophic epidermolysis bullosa. *J Biol Chem.* 2000; 275:24429-24435.
 116. Mavilio F, Pellegrini G, Ferrari S, *et al.* Correction of junctional epidermolysis bullosa by transplantation of genetically modified epidermal stem cells. *Nat Med.* 2006; 12:1397-1402.
 117. De Rosa L, Carulli S, Cocchiarella F, Quaglino D, Enzo E, Franchini E, Giannetti A, De Santis G, Recchia A, Pellegrini G, De Luca M. Long-term stability and safety of transgenic cultured epidermal stem cells in gene therapy of junctional epidermolysis bullosa. *Stem Cell Reports.* 2013; 2:1-8.
 118. Murauer EM, Koller U, Pellegrini G, De Luca M, Bauer JW. Advances in gene/cell therapy in epidermolysis bullosa. *Keio J Med.* 2015; 64:21-25.
 119. Siprashvili Z, Nguyen NT, Gorell ES, Loutit K, Khuu P, Furukawa LK, Lorenz HP, Leung TH, Keene DR, Rieger KE, Khavari P, Lane AT, Tang JY, Marinkovich MP. Safety and wound outcomes following genetically corrected autologous epidermal grafts in patients with recessive dystrophic epidermolysis bullosa. *JAMA.* 2016; 316:1808-1817.
 120. Ortiz-Urda S, Thyagarajan B, Keene DR, Lin Q, Fang M, Calos MP, Khavari PA. Stable nonviral genetic correction of inherited human skin disease. *Nat Med.* 2002; 8:1166-1170.
 121. Ortiz-Urda S, Lin Q, Green CL, Keene DR, Marinkovich MP, Khavari PA. Injection of genetically engineered fibroblasts corrects regenerated human epidermolysis bullosa skin tissue. *J Clin Invest.* 2003; 111:251-255.
 122. Cutlar L, Zhou D, Hu X, Duarte B, Greiser U, Larcher F, Wang W. A non-viral gene therapy for treatment of recessive dystrophic epidermolysis bullosa. *Exp Dermatol.* 2016; 25:818-820.
 123. Pendaries V, Gasc G, Titeux M, Tonasso L, Mejia JE, Hovnanian A. siRNA-mediated allele-specific inhibition of mutant type VII collagen in dominant dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2012; 132:1741-1743.
 124. Morgan CP, Allen DS, Millington-Ward S, O'Dwyer GE, Palfi A, Farrar GJ. A mutation-independent therapeutic strategy for dominant dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2013; 133:2793-2796.
 125. Turczynski S, Titeux M, Tonasso L, Decha A, Ishida-Yamamoto A, Hovnanian A. Targeted exon skipping restores type VII collagen expression and anchoring fibril formation in an *in vivo* RDEB model. *J Invest Dermatol.* 2016; 136:2387-2395.
 126. Murauer EM, Gache Y, Gratz IK, Klausegger A, Muss W, Gruber C, Meneguzzi G, Hintner H, Bauer JW. Functional correction of type VII collagen expression in dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2011; 131:74-83.
 127. Peking P, Koller U, Hainzl S, Kitzmueller S, Kocher T, Mayr E, Nystrom A, Lener T, Reichelt J, Bauer JW, Murauer EM. A gene gun-mediated nonviral RNA trans-splicing strategy for Col7a1 repair. *Mol Ther Nucleic Acids.* 2016; 5:e287.
 128. Zingman LV, Park S, Olson TM, Alekseev AE, Terzic A. Aminoglycoside-induced translational read-through in disease: Overcoming nonsense mutations by pharmacogenetic therapy. *Clin Pharmacol Ther.* 2007; 81:99-103.

129. Bedwell DM, Kaenjak A, Benos DJ, Bebok Z, Bubien JK, Hong J, Tousson A, Clancy JP, Sorscher EJ. Suppression of a *CFTR* premature stop mutation in a bronchial epithelial cell line. *Nat Med.* 1997; 3:1280-1284.
130. Wagner KR, Hamed S, Hadley DW, Gropman AL, Burstein AH, Escolar DM, Hoffman EP, Fischbeck KH. Gentamicin treatment of Duchenne and Becker muscular dystrophy due to nonsense mutations. *Ann Neurol.* 2001; 49:706-711.
131. Christian M, Cermak T, Doyle EL, Schmidt C, Zhang F, Hummel A, Bogdanove AJ, Voytas DF. Targeting DNA double-strand breaks with TAL effector nucleases. *Genetics.* 2010; 186:757-761.
132. Cermak T, Doyle EL, Christian M, Wang L, Zhang Y, Schmidt C, Baller JA, Somia NV, Bogdanove AJ, Voytas DF. Efficient design and assembly of custom TALEN and other TAL effector-based constructs for DNA targeting. *Nucleic Acids Res.* 2011; 39:e82.
133. Jinek M, Chylinski K, Fonfara I, Hauer M, Doudna JA, Charpentier E. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science.* 2012; 337:816-821.
134. Yamanaka S, Takahashi K. Induction of pluripotent stem cells from mouse fibroblast cultures. *Tanpakushitsu Kakusan Koso.* 2006; 51:2346-2351. (in Japanese)
135. Osborn MJ, Starker CG, McElroy AN, *et al.* TALEN-based gene correction for epidermolysis bullosa. *Mol Ther.* 2013; 21:1151-1159.
136. Bilousova G, Chen J, Roop DR. Differentiation of mouse induced pluripotent stem cells into a multipotent keratinocyte lineage. *J Invest Dermatol.* 2011; 131:857-864.
137. Itoh M, Kiuru M, Cairo MS, Christiano AM. Generation of keratinocytes from normal and recessive dystrophic epidermolysis bullosa-induced pluripotent stem cells. *Proc Natl Acad Sci U S A.* 2011; 108:8797-8802.
138. Tolar J, Xia L, Riddle MJ, Lees CJ, Eide CR, McElmurry RT, Titeux M, Osborn MJ, Lund TC, Hovnanian A, Wagner JE, Blazar BR. Induced pluripotent stem cells from individuals with recessive dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2011; 131:848-856.
139. Woodley DT, Wang X, Amir M, Hwang B, Remington J, Hou Y, Uitto J, Keene D, Chen M. Intravenously injected recombinant human type VII collagen homes to skin wounds and restores skin integrity of dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2013; 133:1910-1913.
140. Wang X, Ghasri P, Amir M, Hwang B, Hou Y, Khalili M, Lin A, Keene D, Uitto J, Woodley DT, Chen M. Topical application of recombinant type VII collagen incorporates into the dermal-epidermal junction and promotes wound closure. *Mol Ther.* 2013; 21:1335-1344.
141. Hovnanian A. Systemic protein therapy for recessive dystrophic epidermolysis bullosa: How far are we from clinical translation? *J Invest Dermatol.* 2013; 133:1719-1721.
142. Palazzi X, Marchal T, Chabanne L, Spadafora A, Magnol JP, Meneguzzi G. Inherited dystrophic epidermolysis bullosa in inbred dogs: A spontaneous animal model for somatic gene therapy. *J Invest Dermatol.* 2000; 115:135-137.
143. South AP, Uitto J. Type VII collagen replacement therapy in recessive dystrophic epidermolysis bullosa-how much, how often? *J Invest Dermatol.* 2016; 136:1079-1081.
144. Woodley DT, Cogan J, Wang X, Hou Y, Haghghian C, Kudo G, Keene DR, Chen M. *De novo* anti-type VII collagen antibodies in patients with recessive dystrophic epidermolysis bullosa. *J Invest Dermatol.* 2014; 134:1138-1140.

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