Review

The neurobiology of the Prader-Willi phenotype of fragile X syndrome

Zukhrofi Muzar^{1,2,§}, Reymundo Lozano^{3,§,*}, Alexander Kolevzon^{3,c,e,f}, Randi J. Hagerman^{1,*}

Summary Fragile X syndrome (FXS) is the most common inherited cause of intellectual disability and autism, caused by a CGG expansion to greater than 200 repeats in the promoter region of *FMR1* on the bottom of the X chromosome. A subgroup of individuals with FXS experience hyperphagia, lack of satiation after meals and severe obesity, this subgroup is referred to have the Prader-Willi phenotype of FXS. Prader-Willi syndrome is one of the most common genetic severe obesity disorders known and it is caused by the lack of the paternal 15q11-13 region. Affected individuals suffer from hyperphagia, lack of satiation, intellectual disability, and behavioral problems. Children with fragile X syndrome Prader-Willi phenotye and those with Prader Willi syndrome have clinical and molecular similarities reviewed here which will impact new treatment options for both disorders.

Keywords: Fragile X syndrome (FXS), Prader-Willi phenotype, *FMR1* gene, Hyperphagia, Autism, IGF-1, Growth hormone

1. Introduction

Prader-Willi syndrome (PWS) is the most common cause of obesity and intellectual disability, occurring in about 1 in 15,000 in the general population (*1-3*). PWS is characterized by hyperphagia and lack of satiation after meals. Typical physical features include: a round face, narrow palpebral fissures, short stature, small genitalia, and short fingers and toes. Behavioral and cognitive features include mild intellectual disability (ID), poor attention, obsessive behavior particularly

*Address correspondence to:

around food so that food hoarding and food stealing are common, excessive skin picking, and remarkably good ability with puzzles. PWS is caused by an absence of expression of imprinted genes in the paternally derived PWS/Angelman syndrome (AS) region (15q11.2-q13) of chromosome 15 by one of several genetic mechanisms (paternal deletion, maternal uniparental disomy and rarely an imprinting defect) (2).

Fragile X syndrome (FXS) is the most common inherited cause of ID and Autism Spectrum Disorder (ASD) and it is caused by a CGG trinucleotide expansion of over 200 repeats (full mutation) in the 5' region of the fragile X mental retardation 1 gene (*FMR1*) at Xq27.3. This full mutation leads to hypermethylation of *FMR1* and a subsequent lack of transcription and translation, which in turn results in a deficiency of the FMR1 protein (FMRP). The FMRP is an RNA binding protein that mainly negatively regulates the translation hundreds of genes, many of which are critical for synaptic plasticity (4). The prevalence of FXS is 1 in 5,000 males and 1 in 8,000 females (5). The features of FXS include hyperactivity, attentional problems, poor eye contact, hand-flapping, anxiety, hyperarousal and

^{1 a)}Medical Investigation of Neurodevelopmental Disorders MIND Institute, ^{b)}Department of Pediatrics, UC Davis Medical Center, Sacramento, CA, USA;

² Department of Histology, Universitas Muhammadiyah Sumatera Utara (UMSU) Faculty of Medicine, Medan, North Sumatera, Indonesia;

^{3 c)}Seaver Autism Center for Research and Treatment, ^{d)}Departments of Genetics and Genomic Sciences, ^{e)}Psychiatry, and ^{f)}Pediatrics, Icahn School of Medicine at Mount Sinai, New York, NY, USA.

Released online in J-STAGE as advance publication November 14, 2016.

[§]These authors contributed equally to this works.

Dr. Reymundo Lozano, Seaver Autism Center for Research and Treatment, Icahn School of Medicine at Mount Sinai, One Gustave L. Levy Place, Box 1230, New York, NY 10025, USA. E-mail: Reymundo.lozano@mssm.edu

Dr. Randi J. Hagerman, MIND Institute, UC Davis Health System, 2825 50th Street, Sacramento, CA 95817, USA. E-mail: rjhagerman@ucdavis.edu

a lack of habituation to sensory stimuli. The physical features of FXS include a long face, prominent ears, hyperextensible finger joints and flat feet. Males with FXS have macroorchidism during and after puberty with a normal phallus. Although the clinical phenotype is very different between PWS and FXS there is a subgroup of individuals with FXS that develop hyperphagia, obesity, and hypogonadism or delayed puberty. Because this phenotype looks like those with Prader-Willi syndrome, it is described as Fragile X syndrome - Prader-Willi phenotype (FXS-PWP) (Figure 1A and 1B). This phenotype in FXS was first reported by Fryns et al. (1987) and has subsequently been described by others (6-10). This phenotype is not related to a deletion of 15q11-q13 nor due to maternal uniparental disorders, and it occurs in less than 10% of individuals with FXS (11). Although short stature and small fingers and toes can sometimes occur in the FXS-PWP this is not seen in the majority of cases. Nowicki et al. (2007) found lower cytoplasmic interacting FMR1 protein 1 (CYIP1) levels in the blood of patients with the FXS-PWP compared to those with FXS without the PWP (6). They also found a higher rate of ASD in those with the FXS-PWP compared to FXS alone (6).

2. Neurobiology of FXS with PWP

Patients with the FXS-PWP have hyperphagia that arises in childhood (6), as is the case in PWS. Even though there is no clear molecular explanation underlying hyperphagia in FXS, it is hypothesized that it arises from dysregulation of gamma-aminobutyric acid (GABA) system in the hypothalamus. The lateral hypothalamus (LH) is a critical modulator of feeding (12,13). A previous study by Jennings et al. demonstrated that GABAergic (Vgat-expressing) neurons in the LH are responsible for producing appetitive and consummatory behaviors (14). FXS animal models have lowered GABA subunit receptors, synthesis of GABA, GABAergic input to many regions of the brain, and increased catabolism of GABA (15,16). Similarly, cerebral GABAA receptor expression is reduced in several brain regions of subjects with PWS (17).

There is a high rate of ASD (7 of 13, 54%) in the patients with FXS-PWP (6). This may be related to the reduction mRNA levels of the CYFIP1, which was found to be two to fourfold lower in the patients with FXS-PWP compared to individuals without FXS and patients with FXS without the PWP (6). CYFIP1 is localized to the critical region for PWS at 15q between breakpoint 1 and 2 (18). FMRP binds to CYFIP1 (19) in the execution of its role as a transporter and regulator of translation of mRNAs (20-22). CYFIP1 expression levels are vital for dendritic arborization and neuronal morphological complexity (23). Neurons from CYFIP1 haploinsufficient animals have smaller and less complex dendritic branching both *in vitro* and *in vivo*



Figure 1. 11 year old with FXS and PWP, showing severe obesity, small hands and mild facial dysmorphic features

(23). Another reason could be due to dysregulation of oxytocin (OT) and arginine vasopressin (AVP) in the brain. The dysregulation of the OT system in animals and humans is linked with marked deficits in social behavior and anxiety (24). There is a scarcity of OT producing neurons in the paraventricular nucleus of the hypothalamus (PVN) in individuals with PWS (25) and in the *Fmr1* KO (knock-out) mice (24). OT administration increases trust and diminishes disruptive behavior in individuals with PWS (26); in those with FXS OT therapy improves social anxiety (27).

3. Treatment

In the treatment of autism, use of intensive early behavioral intervention, such as the Early Start Denver Model (ESDM) has been shown to improve developmental and social outcome in addition to normalization of the EEG abnormalities compared to those treated with community behavioral interventions (28). Such intervention is also recommended in young children with FXS both with and without autism or the PWP (29,30). A variety of medication use can be helpful for those with FXS both with and without the PWP including; stimulants which can help with attention and appetite; Selective Serotonin Reuptake Inhibitors (SSRIs), such as sertraline for anxiety (31); and atypical antipsychotics, such as aripiprazole to stabilize mood, improve autism, aggression and/or tantrums (6). However, aripiprazole can cause an increase in weight gain and particularly for those with a CYP2D6 polymorphism that can slow down the metabolism of aripiprazole (32).

There are a variety of new-targeted treatments that have been studied in those with FXS. To enhance the GABA deficits in FXS, the GABAB agonist, arbaclofen, showed initial benefit for those with ASD or low sociability (33). However, the subsequent phase 3 trial in children and adolescents and adults did



Figure 2. Suggested mechanism of action of GH. GH stimulates the liver to release IGF-1, which activates CREB, and this increases FMRP in neurons. The increase of FMRP leads to increased GABAergic input to some areas of the brain.

not demonstrate efficacy (34). The GABAA agonist ganaxolone is being studied in children 6 to 17 years old at the MIND Institute at UC Davis Medical Center and in Belgium utilizing the same protocol (https://www. clinicaltrials.gov [NCT]: ID number NCT01725152). Results from this study are expected before of the end of this year. The mGluR5 antagonists developed by Roche and Novartis have not demonstrated efficacy in adult and childhood studies, but further studies are planned for an mGluR5 antagonist. Minocycline has been studied in children with FXS and has demonstrated some efficacy so it is often utilized clinically (35,36). Other trials included at multiple centers included metadoxine for improving attention and focus and the IGF-1 analogue developed by Neuren for the treatment of behavioral problems. However, those with the FXS-PWP of are not typically included in such clinical trials because their level of obesity is beyond what is acceptable for inclusion in these trials. Therefore, there are no studies of the treatment in those with FXS-PWP.

In contrast, human growth hormone (GH) has been the panacea for treatment of PWS over the last decade (37,38). Most individuals with PWS, but not all, are deficient in GH and studies that were initially focused on improving growth have not only demonstrated this effect, but also improvements in metabolism, body composition, behavior and cognition (38-44). However, GH therapy on occasion can be associated with significant side effects, such as the stimulation of adenoid tissue leading to obstructive sleep apnea so it should be used carefully in the treatment of PWS. GH can also promote the growth of some malignant tumors so such a history is a contraindication for GH therapy. Since GH therapy has been so beneficial to those with PWS and because there are remarkable similarities between PWS and the FXS-PWP, it is possible that those with the FXS-PWP will benefit from GH therapy.

The observed benefits of the IGF-1 analogue in the KO mouse FXS model support the likelihood that GH therapy should be beneficial in the FXS-PWP. GH stimulates the release of IGF-1 by the liver and this may be the mechanism for the benefit of GH therapy. IGF-1 enhances GABA activity that is deficient in FXS both with and without the PWP (Figure 2)(45). In addition, the GABAB receptor-mediated transactivation of IGF-1 receptors leads to cAMP response elementbinding protein (CREB) activation which in turn binds to FMR1 and increases FMRP levels (46). Therefore, GH in FXS-PWP could stimulate the release of IGF-1 to enhance the GABA system, and increase the residual expression of FMR1 particularly in those who are mosaic or partially unmethylated. Further studies are necessary to determine the molecular benefits of IGF-1 analogues and GH in FXS.

In general, metabolic anomalies are suspected in FXS because about 30% are obese (47). Metabolic anomalies including increased glucose uptake and excess protein synthesis in the brain have been reported in *Fmr1* KO mice, while in the fly (*dfmr1*), it has been shown that FMRP is required during brain development and may function in neuroblast reactivation by regulating an output of the insulin signaling pathway (48-52). Metabolic profiling in the Fmr1 KO mice also revealed profound consequences in brain metabolism, which in turn lead to alterations in the metabolic response, along with anomalies in other physiological processes and behaviors (53). Using Drosophila as a model of FXS it has been shown that the dfmr1 has elevated levels of drosophila insulin-like peptide 2 (Dilp2) in the insulin-producing cells which result in elevated insulin-signaling via the PI3K/Akt/ mTOR pathway (54). It is also known that increased insulin-signaling leads to defects in the circadian output pathway and in short and long-term memory



Figure 3. Mechanism of action of metformin. Metformin decreases protein synthesis and insulin signaling (IS) via the AMPK/Akt/mTOR pathway, it also inhibits the lipid and sterol biosynthetic pathways

deficits. Interestingly, pharmacological restoration using metformin, rescued memory deficits in the dfmr1 (54). Metformin is known to decrease body mass index (BMI) and to prevent cognitive deficits in individuals with diabetes (55-59). Metformin decreases protein synthesis and insulin-signaling via the AMPK/Akt/ mTOR pathway, it also inhibits the lipid and sterol biosynthetic pathways (60-63) (Figure 3). Therefore, metformin in FXS may decrease insulin-signaling and restore the circadian output pathway and in turn have positive effects on memory and sleep. A pilot, open-label study of response to metformin in 21 children with PWS and six with early morbid obesity (EMO) showed significant improvements in foodrelated distress, anxiety, and ability to be redirected away from food. Within the PWS group, responders to metformin had higher 2-hour glucose levels on oral glucose tolerance test and higher fasting insulin levels. Additionally, parents of 5/13 individuals with PWS and 5/6 with EMO reported recognition of satiety (64). Further studies are necessary to determine the safety and efficacy of metformin in FXS, PWS and FXS-PWP.

4. Future directions for research

Understanding the similarities and differences between PWS and the PWP of FXS will lead to new treatments perhaps for both disorders. Since *CYFIP1* is down-regulated in the PWP and because the clinical phenotypes are so similar across both disorders, it is likely that epigenetic changes or methylation differences may be down-regulating other genes in the 15 q 11-13 region in those with the PWP. Further studies are warranted to determine whether the reduced expression of multiple genes in PWS also occur in FXS-PWP. These genes include, *MKRN3* (65), *MAGEL2* (66), *MAGED1* (67), *NECDIN* (68,69) and *SNURF-SNRPRN* (70). To understand more about hyperphagia phenotype in FXS-PWP, studies regarding the GABA network in the brain are required. In addition, as chronic hyperghrelinaemia promotes hyperphagia in PWS (71), it would be interesting to see ghrelin levels in patients with the FXS-PWP.

A variety of new treatments are currently being studied in PWS and reviewed in Miller *et al.* (2015). Although GH treatment has many beneficial effects, it does not significantly help the hyperphagia. Some of the new medications that are being studied in PWS include Diazoxide, a potent K⁺-ATP channel agonist that hyperpolarizes hypothalamic neurons whose activity is impaired by a defective leptin signaling pathway in PWS (NCT02-34071); AZP-573, an unacylated ghrelin analog; Exanatide/Liraglutide, glucagon-like peptide 1 (GLP-1) receptor agonist which can suppress appetite and reduce weight in PWS and obese patients (NCT014448981/ NCT01542242). Clearly the study of those with FXS- PWP will determine whether they have the potential to benefit from some of these new trials that have occurred in PWS. The future of treatment of both disorders looks bright with the advent of targeted treatments based on the neurobiological studies of both disorders.

Acknowledgements

This work was supported by grants from the National Fragile X Foundation, NICHD grant HD036071, the Department of Defense PR101054, the MIND Institute Intellectual and Developmental Disabilities Research Center U54 HD079125, the Health and Human Services Administration for Developmental Disabilities grant 90DD0596 and by the Seaver Foundation; Dr. Lozano is a Seaver Faculty, Friedman Brain Institute and NIH faculty Scholar.

References

- Butler MG, Lee PDK, Whitman BY, editors. Management of Prader-Willi Syndrome. New York, NY: Springer New York; 2006. http://link.springer. com/10.1007/978-0-387-33536-0 (accessed September 26, 2016).
- Cassidy SB, Schwartz S, Miller JL, Driscoll DJ. Prader-Willi syndrome. Genet Med. 2012; 14:10-26.
- 3. Butler MG. Prader-Willi Syndrome: Obesity due to genomic imprinting. Curr Genomics. 2011; 12:204-215.
- Fernández E, Rajan N, Bagni C. The FMRP regulon: From targets to disease convergence. Front Neurosci. 2013; 7:191.
- Tassone F, Iong KP, Tong T-H, Lo J, Gane LW, Berry-Kravis E, Nguyen D, Mu LY, Laffin J, Bailey DB, Hagerman RJ. *FMR1* CGG allele size and prevalence ascertained through newborn screening in the United States. Genome Med. 2012; 4:100.
- Nowicki ST, Tassone F, Ono MY, Ferranti J, Croquette MF, Goodlin-Jones B, Hagerman RJ. The Prader-Willi phenotype of fragile X syndrome. J Dev Behav Pediatr. 2007; 28:133-138.
- Fryns JP, Haspeslagh M, Dereymaeker AM, Volcke P, Van den Berghe H. A peculiar subphenotype in the fra(X) syndrome: Extreme obesity-short stature-stubby hands and feet-diffuse hyperpigmentation. Further evidence of disturbed hypothalamic function in the fra(X) syndrome? Clin Genet. 1987; 32:388-392.
- Schrander-Stumpel C, Gerver WJ, Meyer H, Engelen J, Mulder H, Fryns JP. Prader-Willi-like phenotype in fragile X syndrome. Clin Genet. 1994; 45:175-180.
- de Vries BB, Fryns JP, Butler MG, Canziani F, Wesbyvan Swaay E, van Hemel JO, Oostra BA, Halley DJ, Niermeijer MF. Clinical and molecular studies in fragile X patients with a Prader-Willi-like phenotype. J Med Genet. 1993; 30:761-766.
- De Vries BB, Niermeijer MF. The Prader-Willi-like phenotype in fragile X patients: A designation facilitating clinical (and molecular) differential diagnosis. J Med Genet. 1994; 31:820.
- 11. McLennan Y, Polussa J, Tassone F, Hagerman R. Fragile x syndrome. Curr Genomics. 2011; 12:216-224.
- 12. HOEBEL BG, TEITELBAUM P. Hypothalamic control

of feeding and self-stimulation. Science. 1962; 135:375-377.

- Berthoud H-R, Münzberg H. The lateral hypothalamus as integrator of metabolic and environmental needs: From electrical self-stimulation to opto-genetics. Physiol Behav. 2011; 104:29-39.
- Jennings JH, Ung RL, Resendez SL, Stamatakis AM, Taylor JG, Huang J, Veleta K, Kantak PA, Aita M, Shilling-Scrivo K, Ramakrishnan C, Deisseroth K, Otte S, Stuber GD. Visualizing hypothalamic network dynamics for appetitive and consummatory behaviors. Cell. 2015; 160:516-527.
- Berry-Kravis E, Knox A, Hervey C. Targeted treatments for fragile X syndrome. J Neurodev Disord. 2011; 3:193-210.
- Hagerman R, Lozano R, Hare E. Modulation of the GABAergic pathway for the treatment of fragile X syndrome. Neuropsychiatr Dis Treat. 2014; 10:1769-1779.
- Lucignani G, Panzacchi A, Bosio L, Moresco RM, Ravasi L, Coppa I, Chiumello G, Frey K, Koeppe R, Fazio F. GABA_A receptor abnormalities in Prader-Willi syndrome assessed with positron emission tomography and [¹¹C]flumazenil. Neuroimage. 2004; 22:22-28.
- 18. Chai J-H, Locke DP, Greally JM, Knoll JHM, Ohta T, Dunai J, Yavor A, Eichler EE, Nicholls RD. Identification of four highly conserved genes between breakpoint hotspots BP1 and BP2 of the Prader-Willi/Angelman syndromes deletion region that have undergone evolutionary transposition mediated by flanking duplicons. Am J Hum Genet. 2003; 73:898-925.
- Schenck A, Bardoni B, Moro A, Bagni C, Mandel JL. A highly conserved protein family interacting with the fragile X mental retardation protein (FMRP) and displaying selective interactions with FMRP-related proteins FXR1P and FXR2P. Proc Natl Acad Sci U S A. 2001; 98:8844-8849.
- Bardoni B, Mandel J-L. Advances in understanding of fragile X pathogenesis and FMRP function, and in identification of X linked mental retardation genes. Curr Opin Genet Dev. 2002; 12:284-293.
- Schenck A, Bardoni B, Langmann C, Harden N, Mandel JL, Giangrande A. CYFIP/Sra-1 controls neuronal connectivity in Drosophila and links the Rac1 GTPase pathway to the fragile X protein. Neuron. 2003; 38:887-898.
- 22. De Rubeis S, Pasciuto E, Li KW, *et al.* CYFIP1 coordinates mRNA translation and cytoskeleton remodeling to ensure proper dendritic spine formation. Neuron. 2013; 79:1169-1182.
- 23. Pathania M, Davenport EC, Muir J, Sheehan DF, López-Doménech G, Kittler JT. The autism and schizophrenia associated gene *CYFIP1* is critical for the maintenance of dendritic complexity and the stabilization of mature spines. Transl Psychiatry. 2014; 4:e374.
- Francis SM, Sagar A, Levin-Decanini T, Liu W, Carter CS, Jacob S. Oxytocin and vasopressin systems in genetic syndromes and neurodevelopmental disorders. Brain Res. 2014; 1580:199-218.
- Swaab DF, Purba JS, Hofman MA. Alterations in the hypothalamic paraventricular nucleus and its oxytocin neurons (putative satiety cells) in Prader-Willi syndrome: A study of five cases. J Clin Endocrinol Metab. 1995; 80:573-579.
- 26. Tauber M, Mantoulan C, Copet P, Jauregui J, Demeer

G, Diene G, Rogé B, Laurier V, Ehlinger V, Arnaud C, Molinas C, Thuilleaux D. Oxytocin may be useful to increase trust in others and decrease disruptive behaviours in patients with Prader-Willi syndrome: A randomised placebo-controlled trial in 24 patients. Orphanet J Rare Dis. 2011; 6:47.

- Hall SS, Lightbody AA, McCarthy BE, Parker KJ, Reiss AL. Effects of intranasal oxytocin on social anxiety in males with fragile X syndrome. Psychoneuroendocrinology. 2012; 37:509-518.
- Dawson G, Jones EJH, Merkle K, Venema K, Lowy R, Faja S, Kamara D, Murias M, Greenson J, Winter J, Smith M, Rogers SJ, Webb SJ. Early behavioral intervention is associated with normalized brain activity in young children with autism. J Am Acad Child Adolesc Psychiatry. 2012; 51:1150-1159.
- 29. Tolmie J. Fragile X syndrome diagnosis, treatment and research. J Med Genet. 2002; 39:783.
- Hagerman RJ, Berry-Kravis E, Kaufmann WE, Ono MY, Tartaglia N, Lachiewicz A, Kronk R, Delahunty C, Hessl D, Visootsak J, Picker J, Gane L, Tranfaglia M. Advances in the treatment of fragile X syndrome. Pediatrics. 2009; 123:378-390.
- 31. Indah Winarni T, Chonchaiya W, Adams E, Au J, Mu Y, Rivera SM, Nguyen DV, Hagerman RJ. Sertraline may improve language developmental trajectory in young children with fragile x syndrome: A retrospective chart review. Autism Res Treat. 2012; 2012:104317.
- Nussbaum LA, Dumitraşcu V, Tudor A, Grădinaru R, Andreescu N, Puiu M. Molecular study of weight gain related to atypical antipsychotics: Clinical implications of the CYP2D6 genotype. Rom J Morphol Embryol. 2014; 55:877-884.
- 33. Berry-Kravis EM, Hessl D, Rathmell B, Zarevics P, Cherubini M, Walton-Bowen K, Mu Y, Nguyen DV, Gonzalez-Heydrich J, Wang PP, Carpenter RL, Bear MF, Hagerman RJ. Effects of STX209 (arbaclofen) on neurobehavioral function in children and adults with fragile X syndrome: A randomized, controlled, phase 2 trial. Sci Transl Med. 2012; 4:152ra127.
- Berry-Kravis E. Arbaclofen In Fragile X Syndrome: Results Of Phase 3 Trials and FXCRC Analysis Of Arbaclofen Respons. In: National Fragile X Foundation, 14th International Fragile X Conference, Orange County, CA. USA, 2014.
- 35. Leigh MJS, Nguyen DV, Mu Y, Winarni TI, Schneider A, Chechi T, Polussa J, Doucet P, Tassone F, Rivera SM, Hessl D, Hagerman RJ. A randomized doubleblind, placebo-controlled trial of minocycline in children and adolescents with fragile X syndrome. J Dev Behav Pediatr. 2013; 34:147-155.
- 36. Lozano R, Hagerman RJ. Treatment of Neurodevelopmental Disorders. In: Hagerman RJ, Hendren RL, editors. Treatment of neurodevelopmental disorders: Targeting neurobiological mechanisms. Oxford University Press, Oxford, UK, 2014.
- 37. Deal CL, Tony M, Höybye C, Allen DB, Tauber M, Christiansen JS, 2011 Growth hormone in prader-Willi Syndrome clinical care guidelines workshop participants. Growth hormone research society workshop summary: Consensus guidelines for recombinant human growth hormone therapy in Prader-Willi syndrome. J Clin Endocrinol Metab. 2013; 98:E1072-E1087.
- Miller J, Strong T, Heinemann J. Medication Trials for Hyperphagia and Food-Related Behaviors in Prader-

Willi Syndrome. Diseases. 2015; 3:78-85.

- Angulo MA, Castro-Magana M, Lamerson M, Arguello R, Accacha S, Khan A. Final adult height in children with Prader-Willi syndrome with and without human growth hormone treatment. Am J Med Genet A. 2007; 143A:1456-1461.
- 40. Carrel AL, Myers SE, Whitman BY, Eickhoff J, Allen DB. Long-term growth hormone therapy changes the natural history of body composition and motor function in children with prader-willi syndrome. J Clin Endocrinol Metab. 2010; 95:1131-1136.
- Butler MG, Smith BK, Lee J, Gibson C, Schmoll C, Moore WV, Donnelly JE. Effects of growth hormone treatment in adults with Prader-Willi syndrome. Growth Horm IGF Res. 2013; 23:81-87.
- 42. Siemensma EPC, Tummers-de Lind van Wijngaarden RFA, Festen DA, *et al.* Beneficial effects of growth hormone treatment on cognition in children with Prader-Willi syndrome: A randomized controlled trial and longitudinal study. J Clin Endocrinol Metab. 2012; 97:2307-2314.
- Sanchez-Ortiga R, Klibanski A, Tritos NA. Effects of recombinant human growth hormone therapy in adults with Prader-Willi syndrome: A meta-analysis. Clin Endocrinol (Oxf). 2012; 77:86-93.
- 44. Mogul HR, Lee PDK, Whitman BY, Zipf WB, Frey M, Myers S, Cahan M, Pinyerd B, Southren AL. Growth hormone treatment of adults with Prader-Willi syndrome and growth hormone deficiency improves lean body mass, fractional body fat, and serum triiodothyronine without glucose impairment: Results from the United States multicenter trial. J Clin Endocrinol Metab. 2008; 93:1238-1245.
- Maya-Vetencourt JF, Baroncelli L, Viegi A, Tiraboschi E, Castren E, Cattaneo A, Maffei L. IGF-1 restores visual cortex plasticity in adult life by reducing local GABA levels. Neural Plast. 2012; 2012:250421.
- 46. Zhang W, Xu C, Tu H, Wang Y, Sun Q, Hu P, Rondard P, Liu J. GABA_B receptor upregulates fragile X mental retardation protein expression in neurons. Sci Rep. 2015; 5:10468.
- 47. McLennan Y, Polussa J, Tassone F, Hagerman R. Fragile x syndrome. Curr Genomics. 2011; 12:216-224.
- Davis RL. Olfactory memory formation in Drosophila: From molecular to systems neuroscience. Annu Rev Neurosci. 2005; 28:275-302.
- Griffith LC, Ejima A. Multimodal sensory integration of courtship stimulating cues in Drosophila melanogaster. Ann N Y Acad Sci. 2009; 1170:394-398.
- Joiner MlA null, Griffith LC. CaM kinase II and visual input modulate memory formation in the neuronal circuit controlling courtship conditioning. J Neurosci Off J Soc Neurosci. 1997; 17:9384-9391.
- McGuire SE, Le PT, Osborn AJ, Matsumoto K, Davis RL. Spatiotemporal rescue of memory dysfunction in Drosophila. Science. 2003; 302:1765-1768.
- Nässel DR, Kubrak OI, Liu Y, Luo J, Lushchak O V. Factors that regulate insulin producing cells and their output in Drosophila. Front Physiol. 2013; 4:252.
- 53. Qin M, Kang J, Smith CB. Increased rates of cerebral glucose metabolism in a mouse model of fragile X mental retardation. Proc Natl Acad Sci U S A. 2002; 99:15758-15763.
- 54. Monyak RE, Emerson D, Schoenfeld BP, Zheng X, Chambers DB, Rosenfelt C, Langer S, Hinchey P, Choi

CH, McDonald TV, Bolduc FV, Sehgal A, McBride SM, Jongens TA. Insulin signaling misregulation underlies circadian and cognitive deficits in a Drosophila fragile X model. Mol Psychiatry. 2016; doi: 10.1038/mp.2016.51.

- 55. McDonagh MS, Selph S, Ozpinar A, Foley C. Systematic review of the benefits and risks of metformin in treating obesity in children aged 18 years and younger. JAMA Pediatr. 2014; 168:178-184.
- Ying MA, Maruschak N, Mansur R, Carvalho AF, Cha DS, McIntyre RS. Metformin: Repurposing opportunities for cognitive and mood dysfunction. CNS Neurol Disord Drug Targets. 2014; 13:1836-1845.
- Ng TP, Feng L, Yap KB, Lee TS, Tan CH, Winblad B. Long-term metformin usage and cognitive function among older adults with diabetes. J Alzheimer's Dis JAD. 2014; 41:61-68.
- Sun W, Zeng C, Liao L, Chen J, Wang Y. Comparison of acarbose and metformin therapy in newly diagnosed type 2 diabetic patients with overweight and/or obesity. Curr Med Res Opin. 2016; 32:1389-1396.
- Roumie CL, Greevy RA, Grijalva CG, Hung AM, Liu X, Griffin MR. Diabetes treatment intensification and associated changes in HbA1c and body mass index: A cohort study. BMC Endocr Disord. 2016; 16:32.
- 60. Jin J, Gu H, Anders NM, Ren T, Jiang M, Tao M, Peng Q, Rudek MA, Duan W. Metformin protects cells from mutant huntingtin toxicity through activation of AMPK and modulation of mitochondrial dynamics. Neuromolecular Med. 2016; doi: 10.1007/s12017-016-8412-z
- Luo T, Nocon A, Fry J, Sherban A, Rui X, Jiang B, Xu XJ, Han J, Yan Y, Yang Q, Li Q, Zang M. AMPK activation by metformin suppresses abnormal adipose tissue extracellular matrix remodeling and ameliorates insulin resistance in obesity. Diabetes. 2016; 65:2295-2310.
- Anabtawi A, Miles JM. Metformin: Non-glycemic effects and potential novel indications. Endocr Pract. 2016; doi: 10.4158/EP151145.RAR
- 63. Rattan R, Ali Fehmi R, Munkarah A. Metformin: An emerging new therapeutic option for targeting cancer stem cells and metastasis. J Oncol. 2012; 2012:928127.

- Miller JL, Linville TD, Dykens EM. Effects of metformin in children and adolescents with Prader-Willi syndrome and early-onset morbid obesity: A pilot study. J Pediatr Endocrinol Metab JPEM. 2014; 27:23-29.
- Kanber D, Giltay J, Wieczorek D, Zogel C, Hochstenbach R, Caliebe A, Kuechler A, Horsthemke B, Buiting K. A paternal deletion of *MKRN3*, *MAGEL2* and *NDN* does not result in Prader-Willi syndrome. Eur J Hum Genet. 2009; 17:582-590.
- 66. Boccaccio I, Glatt-Deeley H, Watrin F, Roëckel N, Lalande M, Muscatelli F. The human *MAGEL2* gene and its mouse homologue are paternally expressed and mapped to the Prader-Willi region. Hum Mol Genet. 1999; 8:2497-2505.
- 67. Dombret C, Nguyen T, Schakman O, Michaud JL, Hardin-Pouzet H, Bertrand MJ, De Backer O. Loss of *Maged1* results in obesity, deficits of social interactions, impaired sexual behavior and severe alteration of mature oxytocin production in the hypothalamus. Hum Mol Genet. 2012; 21:4703-4717.
- 68. Muscatelli F, Abrous DN, Massacrier A, Boccaccio I, Le Moal M, Cau P, Cremer H. Disruption of the mouse Necdin gene results in hypothalamic and behavioral alterations reminiscent of the human Prader-Willi syndrome. Hum Mol Genet. 2000; 9:3101-3110.
- Lavi-Itzkovitz A, Tcherpakov M, Levy Z, Itzkovitz S, Muscatelli F, Fainzilber M. Functional Consequences of Necdin Nucleocytoplasmic Localization. PLoS One. 2012; 7:e33786.
- 70. Tsai TF, Jiang YH, Bressler J, Armstrong D, Beaudet AL. Paternal deletion from Snrpn to Ube3a in the mouse causes hypotonia, growth retardation and partial lethality and provides evidence for a gene contributing to Prader-Willi syndrome. Hum Mol Genet. 1999; 8:1357-1364.
- Haqq AM, Grambow SC, Muehlbauer M, Newgard CB, Svetkey LP, Carrel AL, Yanovski JA, Purnell JQ, Freemark M. Ghrelin concentrations in Prader-Willi syndrome (PWS) infants and children: Changes during development. Clin Endocrinol (Oxf). 2008; 69:911-920.

(Received October 9, 2016; Revised November 7, 2016; Accepted November 8, 2016)