

An up-date on novel molecular targets in testicular germ cell tumors subtypes

Paolo Chieffi*

Dipartimento di Psicologia, Università della Campania, Caserta, Italy.

Summary

Testicular germ cell tumors (TGCTs) are the most frequent solid malignant tumors in men 20-34 years of age and the most frequent cause of death from solid tumors in this age group. In addition, the incidence of these tumors has significantly increased over the last few decades. Testicular germ cell tumors are classified into seminoma and nonseminoma germ cell tumors (NSGCTs). NSGCTs can be further divided into embryonal carcinoma, Teratoma, yolk sac tumor, and choriocarcinoma. There are noteworthy differences about therapy and prognosis of seminomas and nonseminoma germ cell tumors, even though both share characteristics of the primordial germ cells (PGCs). Many discovered biomarkers including HMGA1, GPR30, Aurora-B, estrogen receptor β , and others have given further advantage to discriminate between histological subgroups and could represent useful molecular therapeutic targets.

Keywords: Testicular germ cells tumors, seminomas, Aurora B, GPR30, PATZ1, HMGA

Testicular germ cell tumors (TGCTs) have the highest incidence among young men (between 20 and 34 years of age) of solid tumors, and their incidence has significantly increased over the last few decades. About 90% of TGCTs are successfully treated with cisplatin-based chemotherapy. However, this kind of therapy raises the possibility of developing secondary cancers and cardiovascular disease. TGCTs are classified into two principal groups: Germ Cell Neoplasias *In Situ* (GCNIS) that are Seminoma and Nonseminoma (NSE), and spermatocytic tumors that are not GCNIS. NSE tumors encompass embryonal carcinoma, choriocarcinoma, Yolk Sac Tumors (YSTs) and teratomas. TGCTs may develop from a non-invasive type of tumor called carcinoma *in situ* (CIS): Microscope analysis reveals abnormal cells even though they are still confined inside the membrane of the seminiferous tubules (1-9).

A significant increase in TGCTs incidence occurred in the last few decades, probably due to altered

environmental factors that significantly contribute to disease onset. For instance, although the biological mechanisms are still unclear, evidence suggests that the risk of developing TGCTs is associated with maternal smoking during pregnancy, adult height, biomass index, diet rich in cheese, pesticide exposure, and others. Among the risk factors involved in the onset of disease: age, cryptorchidism, family history of testicular cancer, Klinefelter's syndrome, personal history of testicular cancer, congenital abnormalities and infertility. Cryptorchidism is the major risk factor associated with germ cell tumors: it deals with undescended testicle into the scrotum, which remains in the abdomen or groin, thus the risk of developing the disease does not change even after surgery to move the testicle into the scrotum. Remarkably, it is still debatable whether the exposure to some nonsteroidal estrogens during pregnancy, such as diethylstilbestrol (DES) may increase the risk of developing TGCTs. Despite that this divergent evidence confirms the important role played by some environmental factors in TGCTs, etiology has been clearly suggested by migration studies. Consistently, Sweden has an incidence of TGCTs about twice that of Finland and, although first generation migrants from Finland to Sweden show no increased risk, second generation males born to the migrant parents in Sweden present an increased frequency (10,11).

Released online in J-STAGE as advance publication May 29, 2019.

*Address correspondence to:

Dr. Paolo Chieffi, Dipartimento di Psicologia, Università della Campania, Viale Ellittico, 3181100 Caserta, Italy.
E-mail: paolo.chieffi@unicampania.it

Numerous new biomarkers have been found to discriminate TGCTs subtypes, standing for innovative molecular therapeutic targets. High-mobility group proteins A1 (HMGA1) and A2 (HMGA2) act as powerful diagnostic markers (12-15). Really, these two proteins are diversely expressed in TGCTs in comparison with the stage of tumor differentiation (12,13). For example, HMGA1 binds to other proteins, such as RNF4 (16,17) and PATZ1, which are engaged in transcriptional control and have been demonstrated to be overexpressed and delocalized in human testicular seminomas (18). Currently, we have shown that in human testicular seminomas Estrogen Receptor β (ER β) expression is strongly down regulated and this down regulation is associated with delocalization of both PATZ1 and HMGA1 transcriptional factors, on the contrary, in normal germ cells, PATZ1 binds to ER β (19,20).

The serine/threonine kinase NEK2 is a key regulator of centrosome separation and bipolar spindle formation during mitosis and chromatin condensation during meiosis. It controls centrosome separation (essential for the formation of bipolar spindles and high-fidelity chromosome separation) through the phosphorylation of proteins such as CEP250, CROCC and NINL, causing their dislocation from the centrosomes. Additionally, NEK2 has a major function in chromatin condensation in the first meiotic division by HMGA2 phosphorylation (21). Moreover, the enhancement and the nuclear localization of NEK2 protein has been found in both seminomas and in seminoma cell line (TCam-2) (22,23). Furthermore, recent studies underlined the new splicing factor kinase function of NEK2 (23).

The RNA-binding protein LIN28 is implicated in the maintenance of the pluripotency of embryonic stem cells, and its expression levels are reduced throughout differentiation. In particular, LIN28 regulates the expression of OCT4 through directly binding to its mRNA transcript in mouse embryonic stem cells. Indeed, LIN28 has a pivotal function for reprogramming somatic cells into pluripotent stem cells. Moreover, LIN28 represents a valid diagnostic marker for testicular GCNIS, classical seminomas, embryonal carcinomas, and YSTs (24). In particular, LIN28 is the main YST marker due to the absence of OCT4 (24).

Estrogen signaling is mediated by two nuclear receptors, estrogen receptor α (ER α) and β (ER β), that are estrogen dependent transcription factors. ER α is expressed at high levels in human epididymis and efferent ductules, but not in the testis, whereas ER β is expressed in spermatogonia, spermatocytes, and in early round spermatids in human testis (1,2). The ER β subtype is the principal mediator of estrogen action in promoting germ cell survival and development. After activation, these receptors, in association with a myriad of co-activators and repressors, act as nuclear transcription factors for targeted genes. It has been

well documented in the literature that ER β , which is expressed in normal testicular cells, is instead down regulated in seminomas and embryonal cell carcinomas (1,2). Until recently, the estrogen receptors α (ER α) and estrogen receptors β (ER β) (25-27) have been considered the major physiologic estrogen mediators. Indeed, the G protein-coupled estrogen receptor (GPR30) has proved to have an increasing role in estrogen-mediated signalling in a wide variety of cell types. The critical role of GPR30 in preservation and in development and homeostasis of normal testis is well recognized (28-30). Recent studies show that GPR30 is overexpressed in human spermatogonia, spermatocytes (29), in the TCam-2 cell line and seminomas. Moreover, it has been verified that ER β downregulation correlates with GPR30 overexpression both in human CIS and seminomas; furthermore, it has been demonstrated that 17 β -estradiol produces ERK1/2 activation through GPR30 (31). Many studies are committed to develop novel therapeutic strategies for the treatment of TGCTs blocking neoplastic germ cells through the design of selective GPR30 inhibitors.

The kinase Aurora-B is another valuable marker able to discriminate among the different tumor histotypes; in fact, it is detected in IGCNU, seminomas and embryonal carcinomas, but not in teratomas and YST. Pharmacological inhibition of Aurora B significantly decreases the cell growth in testicular GC1 and TCam2 cell lines (32-35).

Perturbation of miRNAs plays an important role in the establishment and progression of many cancer types, including TGCTs (36). Although different miRNA signatures are associated with histological subtypes of TGCT, very few miRNAs have been found to have a key role in TGCTs. Indeed, Dicer knockout mice show a premature reduction of germ cell numbers and deregulated differentiation of male germ cells (36). Then, Voorhoeve *et al.* showed that miR-372 and miR-373 may overcome p53-mediated arrest of the cell cycle (37). Conversely, miR-372 and miR-373 were absent in TGCT-derived cell lines with mutated p53 or expressed low levels of p53, suggesting that these miRNAs may allow the growth of TGCT escaping the p53 checkpoint of the cell-cycle. In this context, data suggests that miR-372 and miR-373 may act as oncogenes in TGCT through the inhibition of LATS2, a tumor suppressor gene (36). Moreover, the novel identification of circulating miRNAs in body fluids like serum, may represent a valid non-invasive manner to diagnosis and follow disease status. In this regard, it has been reported that miR-371 and miR-372 are specifically increased in serum of germ cell tumor patients. Moreover, many other miRNAs have been proposed to be able to discriminate between different tumor histotypes, confirming the function of the embryonic miR-371 and miR-372 in identifying malignant TGCT (38).

Pseudogenes have long been considered as non-

functional genomic sequences. However, recent evidence suggests that many of them might have some form of biological activity, and the possibility of functionality through a microRNA-mediated pathway (39). Recently, two HMGA1 processed pseudogenes (HMGA1P6 and HMGA1P7) were isolated. In particular, these pseudogenes, competing with HMGA1 for microRNA binding, lead to the upregulation of HMGA1 cellular levels, exerting an oncogenic role (40). In this context, although further experiments are needed, preliminary data show that HMGA1 pseudogenes are differentially overexpressed in TGCT histotypes in comparison with normal testis (seminomas, embryonal carcinomas, mixed form teratomas, and YSTs), suggesting a role of HMGA1 pseudogenes in TGCT carcinogenesis.

The development of human TGCTs is subjected to genetic and environmental factors that have a crucial role in deregulating the normal differentiation process in PGCs. Recently, the increasing number of tumor biomarkers has permitted histological discrimination among the various subgroups. A better comprehension of the molecular pathways through which the TGCTs develop will point out new tools to definitely target cancer cells and will help to defeat intrinsic and acquired chemotherapy resistance. Aurora-B serine-threonine kinases, HMGA and GPR30 inhibitors (41,42) are promising molecules able to selectively target cancer cells, introducing a new scenario for TGCTs treatment in the near future.

References

- Chieffi P. Molecular targets for the treatment of testicular germ cell tumors. *Mini Rev Med Chem.* 2007; 7:755-759.
- Chieffi P, Franco R, Portella G. Molecular and cell biology of testicular germ cell tumors. *Int Rev Cell Mol Biol.* 2009; 278:277-308.
- Oosterhuis, JW, Looijenga LH. Testicular germ-cell tumors in a broader perspective. *Nat Rev Cancer.* 2005; 5:210-222.
- Chieffi P, Chieffi S, Franco R, Sinisi AA. Recent advances in the biology of germ cell tumors: Implications for the diagnosis and treatment. *J Endocrinol Invest.* 2012; 35:1015-1020.
- Chieffi P, Chieffi S. Molecular biomarkers as potential targets for therapeutic strategies in human testicular germ cell tumours: An overview. *J Cell Physiol.* 2013; 228:1641-1646.
- Chieffi P, Chieffi S. An up-date on newly discovered immunohistochemical biomarkers for the diagnosis of human testicular germ cell tumors. *Histol Histopathol.* 2014; 29:999-1006.
- Chieffi P. An overview on predictive biomarkers of testicular germ cell tumors. *J Cell Physiol.* 2017; 232:276-280.
- Chieffi P. Potential new anticancer molecular targets for the treatment of human testicular seminomas. *Mini Rev Med Chem.* 2011; 11:1075-1081.
- Picascia A, Stanzione R, Chieffi P, Kisslinger A, Dikic I, Tramontano D. Pyk2 regulates proliferation and differentiation of prostate cells. *Mol Cell Endocrinol.* 2002; 186:81-87.
- Dieckmann KP, Hartmann JT, Classen J, Diederichs M, Pichlmeier U. Is increased body mass index associated with the incidence of testicular germ cell cancer? *J Cancer Res Clin Oncol.* 2009; 135:731-738.
- Montgomery SM, Granath F, Ehlin A, Sparén P, Ekblom A. Germ-cell testicular cancer in offspring of Finnish immigrants to Sweden. *Cancer Epidemiol Biomarkers Prev.* 2005; 14:280-282.
- Chieffi P, Battista S, Barchi M, Di Agostino S, Pierantoni GM, Fedele M, Chiariotti L, Tramontano D, Fusco A. HMGA1 and HMGA2 protein expression in mouse spermatogenesis. *Oncogene.* 2002; 21:3644-3650.
- Franco R, Esposito F, Fedele M, Liguori G, Pierantoni GM, Botti G, Tramontano D, Fusco A, Chieffi P. Detection of high mobility group proteins A1 and A2 represents a valid diagnostic marker in post-puberal testicular germ cell tumours. *J Pathol.* 2008; 214:58-64.
- Esposito F, Tornincasa M, Chieffi P, De Martino I, Pierantoni GM, Fusco A. High-mobility group A1 proteins regulate p53-mediated transcription of Bcl-2 gene. *Cancer Res.* 2010; 70:5379-5388.
- Chieffi P. New perspective on molecular markers as promising therapeutic targets in germ cell tumors. *Intractable Rare Dis Res.* 2016; 5:136-139.
- Pero R, Lembo F, Di Vizio D, Boccia A, Chieffi P, Fedele M, Pierantoni GM, Rossi P, Iuliano R, Santoro M, Viglietto G, Bruni CB, Fusco A, Chiariotti L. RNF4 is a growth inhibitor expressed in germ cells and lost in human testicular tumors. *Am J Pathol.* 2001; 159:1225-1230.
- Pero R, Lembo F, Chieffi P, Del Pozzo G, Fedele M, Fusco A, Bruni CB, Chiariotti L. Translational regulation of a novel testis-specific RNF4 transcript. *Mol Reprod Dev.* 2003; 66:1-7.
- Fedele M, Franco R, Salvatore G, Paronetto MP, Barbagallo F, Pero R, Chiariotti L, Sette C, Tramontano D, Chieffi G, Fusco A, Chieffi P. PATZ1 gene has a critical role in the spermatogenesis and testicular tumours. *J Pathol.* 2008; 215:39-47.
- Esposito F, Boscia F, Franco R, Tornincasa M, Fusco A, Kitazawa S, Looijenga LH, Chieffi P. Down-regulation of estrogen receptor- β associates with transcriptional coregulator PATZ1 delocalization in human testicular seminomas. *J Pathol.* 2011; 224:110-120.
- Esposito F, Boscia F, Gigantino V, Tornincasa M, Fusco A, Franco R, Chieffi P. The high mobility group A1-estrogen receptor β nuclear interaction is impaired in human testicular seminomas. *J Cell Physiol.* 2012; 227:3749-3755.
- Di Agostino S, Fedele M, Chieffi P, Fusco A, Rossi P, Geremia R, Sette C. Phosphorylation of high-mobility group protein A2 by Nek2 kinase during the first meiotic division in mouse spermatocytes. *Mol Biol Cell.* 2004; 15:1224-1232.
- Barbagallo F, Paronetto MP, Franco R, Chieffi P, Dolci S, Fry AM, Geremia R, Sette C. Increased expression and nuclear localization of the centrosomal kinase Nek2 in human testicular seminomas. *J Pathol.* 2009; 217:431-441.
- Naro C, Barbagallo F, Chieffi P, Bourgeois CF, Paronetto MP, Sette C. The centrosomal kinase NEK2 is a novel splicing factor kinase involved in cell survival. *Nucleic Acids Res.* 2014; 42:3218-3227.

24. Cao D, Allan RW, Cheng L, Peng Y, Guo CC, Dahiya N, Akhi S, Li J. RNA-binding protein LIN28 is a marker for testicular germ cell tumors. *Hum Pathol.* 2011; 42:710-718.
25. Vicini E, Loiarro M, Di Agostino S, Corallini S, Capolunghi F, Carsetti R, Chieffi P, Geremia R, Stefanini M, Sette C. 17- β -estradiol elicits genomic and non-genomic responses in mouse male germ cells. *J Cell Physiol.* 2006; 206:238-245.
26. Staibano S, Franco R, Mezza E, Chieffi P, Sinisi A, Pasquali D, Errico ME, Nappi C, Tremolaterra F, Somma P, Mansueto G, De Rosa G. Loss of oestrogen receptor β , high PCNA and P53 expression and aneuploidy as markers of worse prognosis in ovarian granulosa cell tumours. *Histopathology.* 2003; 43:254-262.
27. Stabile V, Russo M, Chieffi P. 17 β -estradiol induces Akt-1 through estrogen receptor β in the frog (*Rana esculenta*) male germ cells. *Reproduction* 2006; 132:477-484.
28. Chieffi P, Colucci D'Amato GL, Staibano S, Franco R, Tramontano D. Estradiol-induced mitogen-activated protein kinase (extracellular r signal-regulated kinase 1 and 2) activity in the frog (*Rana esculenta*) testis. *J Endocrinol.* 2000; 167:77-84.
29. Franco R, Boscia F, Gigantino V, Marra L, Esposito F, Ferrara D, Pariante P, Botti G, Caraglia M, Minucci S, Chieffi P. GPR30 is over-expressed in post puberal testicular germ cell tumors. *Cancer Biol Ther.* 2011; 11:609-613.
30. Chieffi P, Franco R, Fulgione D, Staibano S. PCNA in the testis of the frog, *Rana esculenta*: A molecular marker of the mitotic testicular epithelium proliferation. *Gen Comp Endocrinol.* 2000; 119: 11-16.
31. Boscia F, Passaro C, Gigantino V, Perdonà S, Franco R, Portella G, Chieffi S, Chieffi P. High levels of GPR30 protein in human testicular carcinoma *in situ* and seminomas correlate with low levels of estrogen receptor- β and indicate a switch in estrogen responsiveness. *J Cell Physiol.* 2015; 230: 1290-1297.
32. Chieffi P, Troncone G, Caleo A, Libertini S, Linardopoulos S, Tramontano D, Portella G. Aurora B expression in normal testis and seminomas. *J Endocrinol.* 2004; 181:263-270.
33. Esposito F, Libertini S, Franco R, Abagnale A, Marra L, Portella G, Chieffi P. Aurora B expression in post-puberal testicular germ cell tumours. *J Cell Physiol.* 2009; 221:435-439.
34. Portella G, Passaro C, Chieffi P. Aurora B: A new prognostic marker and therapeutic target in cancer. *Curr Med Chem.* 2011; 18:482-496.
35. Libertini S, Abagnale A, Passaro C, Botta G, Barbato S, Chieffi P, Portella G. AZD1152 negatively affects the growth of anaplastic thyroid carcinoma cells and enhances the effects of oncolytic virus *dl922-947*. *Endocr Related Cancer.* 2011; 18:129-141.
36. Chieffi P. An overview on new anticancer molecular targets in human testicular germ cell tumors. *Rendiconti Lincei. Scienze Fisiche e Naturali.* 2014; 25: 221-228.
37. Voorhoeve PM, le Sage C, Schrier M, Gillis AJ, Stoop H, Nagel R, Liu YP, van Duijse J, Drost J, Griekspoor A, Zlotorynski E, Yabuta N, De Vita G, Nojima H, Looijenga LH, Agami R. A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in testicular germ cell tumors. *Cell.* 2006; 124:1169-1181.
38. Rijlaarsdam MA, van Agthoven T, Gillis AJ, Patel S, Hayashibara K, Lee KY, Looijenga LH. Identification of known and novel germ cell cancer-specific (embryonic) miRs in serum by high-throughput profiling. *Andrology.* 2015; 3:85-91.
39. Poliseno L, Salmena L, Zhang J, Carver B, Haveman WJ, Pandolfi PP. A coding-independent function of gene and pseudogene mRNAs regulates tumour biology. *Nature.* 2010; 465:1033-1038.
40. Esposito F, De Martino M, Forzati F, Fusco A. HMGA1-pseudogene overexpression contributes to cancer progression. *Cell Cycle.* 2014; 13:3636-3639.
41. Chieffi P, Boscia F. New discovered molecular markers as promising therapeutic targets in germ cell tumors. *Expert Opinion Orphan Drugs.* 2015; 3:1021-1030.
42. Chieffi P, De Martino M, Esposito F. New anti-cancer strategies in testicular germ cell tumors. *Recent Pat Anticancer Drug Discov.* 2019; 14:53-59.

(Received May 2, 2019; Revised May 11, 2019; Accepted May 23, 2019)